

# Population genomic structure in Goodman's mouse lemur reveals long-standing separation of Madagascar's Central Highlands and eastern rainforests

George P. Tiley<sup>1</sup>  | Tobias van Elst<sup>2</sup>  | Helena Teixeira<sup>2</sup> | Dominik Schüßler<sup>3</sup>  |  
 Jordi Salmons<sup>4</sup>  | Marina B. Blanco<sup>5</sup> | José M. Ralison<sup>6</sup> |  
 Blanchard Randrianambinina<sup>7,8</sup> | Rodin M. Rasoloarison<sup>6,7,9</sup> | Amanda R. Stahlke<sup>10</sup>  |  
 Paul A. Hohenlohe<sup>10</sup>  | Lounès Chikhi<sup>4,11</sup>  | Edward E. Louis<sup>12</sup> | Ute Radespiel<sup>2</sup>  |  
 Anne D. Yoder<sup>1</sup> 

<sup>1</sup>Department of Biology, Duke University, Durham, North Carolina, USA

<sup>2</sup>Institute of Zoology, University of Veterinary Medicine Hannover, Hannover, Germany

<sup>3</sup>Research Group Vegetation Ecology and Nature Conservation, Institute of Biology and Chemistry, University of Hildesheim, Hildesheim, Germany

<sup>4</sup>IRD, UMR5174 EDB (Laboratoire Évolution & Diversité Biologique), CNRS, Université Paul Sabatier, Toulouse, France

<sup>5</sup>Duke Lemur Center, Duke University, Durham, North Carolina, USA

<sup>6</sup>Département de Biologie Animale, Université d'Antananarivo, Antananarivo, Madagascar

<sup>7</sup>Group d'Etude et de Recherche sur les Primates de Madagascar (GERP), Antananarivo, Madagascar

<sup>8</sup>Faculté des Sciences, University of Mahajanga, Mahajanga, Madagascar

<sup>9</sup>Behavioral Ecology and Sociobiology Unit, German Primate Centre, Göttingen, Germany

<sup>10</sup>Department of Biological Sciences, Institute for Bioinformatics and Evolutionary Studies, University of Idaho, Moscow, Idaho, USA

<sup>11</sup>Instituto Gulbenkian de Ciência, Oeiras, Portugal

<sup>12</sup>Grewcock Center for Conservation and Research, Omaha's Henry Doorly Zoo and Aquarium, Omaha, Nebraska, USA

## Correspondence

George P. Tiley, Department of Biology,  
 Duke University, Durham, North Carolina,  
 USA.

Email: [g.tiley@kew.org](mailto:g.tiley@kew.org)

## Present address

George P. Tiley, Royal Botanic Gardens  
 Kew, Richmond TW9 3AE, UK

## Funding information

Alexander von Humboldt-Stiftung;  
 Deutsche Forschungsgemeinschaft,  
 Grant/Award Number: Ra 502/23-1;  
 Duke Tropical Conservation Initiative;  
 Fundação para a Ciência e Tecnologia,  
 Grant/Award Number: PTDC-BIA-  
 EVL/30815/2017; International Research  
 Project-Bioinformatics, Ecology, Evolution,  
 Genomics and Behaviour; Investissement  
 d'Avenir, Grant/Award Number: ANR-10-  
 LABX-25-01; John Simon Guggenheim

## Abstract

Madagascar's Central Highlands are largely composed of grasslands, interspersed with patches of forest. The historical perspective was that Madagascar's grasslands had anthropogenic origins, but emerging evidence suggests that grasslands were a component of the pre-human Central Highlands vegetation. Consequently, there is now vigorous debate regarding the extent to which these grasslands have expanded due to anthropogenic pressures. Here, we shed light on the temporal dynamics of Madagascar's vegetative composition by conducting a population genomic investigation of Goodman's mouse lemur (*Microcebus lehilahytsara*; Cheirogaleidae). These small-bodied primates occur both in Madagascar's eastern rainforests and in the Central Highlands, making them a valuable indicator species. Population divergences among forest-dwelling mammals will reflect changes to their habitat, including fragmentation, whereas patterns of post-divergence gene flow can reveal formerly wooded migration corridors. To explore these patterns, we used RADseq data to infer population genetic

George P. Tiley and Tobias van Elst contributed equally to this work.

Memorial Foundation; Laboratoire d'Excellence TULIP, Grant/Award Number: ANR-10-LABX-41 and ANR-11-IDEX-0002-02; North-German Supercomputing Alliance; Primate Action Fund

**Handling Editor:** Kimberly Gilbert

structure, demographic models of post-divergence gene flow, and population size change through time. The results offer evidence that open habitats are an ancient component of the Central Highlands, and that widespread forest fragmentation occurred naturally during a period of decreased precipitation near the last glacial maximum. Models of gene flow suggest that migration across the Central Highlands has been possible from the Pleistocene through the recent Holocene via riparian corridors. Though our findings support the hypothesis that Central Highland grasslands predate human arrival, we also find evidence for human-mediated population declines. This highlights the extent to which species imminently threatened by human-mediated deforestation may already be vulnerable from paleoclimatic conditions.

#### KEYWORDS

Central Highland Savanna, coalescent, fragmentation, historical demography, *Microcebus*, RAD sequencing

## 1 | INTRODUCTION

Madagascar is an iconic biodiversity hotspot with high levels of species diversity and endemism (Myers et al., 2000). The majority of its biodiversity is at extreme risk of extinction due to deforestation and forest fragmentation (Green & Sussman, 1990; Harper et al., 2007; Vieilledent et al., 2018). It is widely accepted that the rapid rate of biodiversity loss is due largely to human activity and agricultural practices (Farris et al., 2017; Styger et al., 2007). Even so, the timing of and degree to which humans transformed and fragmented Madagascar's forests have been points of contention. Much of central Madagascar is now covered with grasslands, traditionally thought to be strictly the product of fires from slash-and-burn agriculture and cattle farming (Gade, 1996; Humbert, 1927; Perrier de la Bâthie, 1921). The best available archaeological evidence places human arrival in Madagascar between 2000 years ago (ka), consistent with human population genomics (Pierron et al., 2017), and 10 ka (Dewar et al., 2013; Dewar & Wright, 1993; Gommery et al., 2011; Hansford et al., 2018; Mitchell, 2019). This timeline coincides with evidence of increased frequency of fire based on isotope records from stalagmites (Railsback et al., 2020) and charcoal records from sediment cores covering the late to mid Holocene (Burney, 1987a; Gasse & Van Campo, 1998; Virah-Sawmy et al., 2010). Pollen abundances from these same sediment cores also imply that the Central Highland vegetation transitioned from woody taxa to grasses during this timeframe, supporting the hypothesis that the grassy dominance of the Central Highlands is the consequence of human activities (reviewed in Burney et al., 2004; Joseph et al., 2021). However, it becomes difficult to disentangle anthropogenic and paleoclimatic effects given that the transition to grasslands probably began during the climatic fluctuations of the Pleistocene (Burney, 1987b; Gasse & Van Campo, 1998), long before the arrival of humans. Changes in plant composition from woody taxa to grasses have also been reported outside of the Central Highlands in southwestern (Burney, 1993) and southeastern (Virah-Sawmy et al., 2009) Madagascar. Taken together, the paleoecological

data imply that a grassy Central Highland vegetation may not strictly be the product of human agricultural practices, but instead, the consequence of global climate fluctuations, perhaps facilitated by the natural processes of fire ecology (Bowman & Franklin, 2005).

In support of this second scenario, ecological and phylogenetic studies of Madagascar's grasses indicate that Central Highland grasslands originated well before the Holocene, with levels of endemism comparable to the grasslands of South Africa (Bond et al., 2008) and tropical east Africa (Vorontsova et al., 2016) where ancient origins are well-supported. And though few  $C_4$  grass lineages such as *Aristida* and *Loudetia* dominate the Central Highlands (Koechlin, 1993; Kull, 2003; Solofondranohatra et al., 2018), diversity among endemic  $C_4$  grass clades would be more consistent with the global diversification of  $C_4$  grasses during the Miocene (e.g., Edwards & Smith, 2010) rather than as a consequence of many independent migration events (Hackel et al., 2018). We are therefore left with a complex model of Pleistocene climate change, perhaps driving grassland expansions, but overlaid with deforestation and fragmentation, probably associated with and intensified by early human colonization. We hypothesize that present-day distributions of forest-dwelling species, their genetic diversity, and connectivity among populations will reflect both ancient climatic change as well as recent human-mediated processes. Here, we perform a population genomic investigation of Goodman's mouse lemur (*Microcebus lehilahytsara*) to test whether signatures of pre- and post-human vegetative change in Madagascar are detectable with extant molecular data and demographic models.

The geographic distribution of *M. lehilahytsara* is uniquely suited for this purpose. It is the only mouse lemur species found in isolated Central Highland forest patches (Andriambeloson et al., 2021; Yoder et al., 2016) as well as in the humid rainforests along the escarpment of Madagascar's east coast (Blanco et al., 2017; Radespiel et al., 2012). It is also found in the rainforests of northeastern Madagascar, as the formerly recognized *M. mittermeieri* was recently recommended as a junior synonym of *M. lehilahytsara* based on both morphological (Schüßler, Blanco, et al., 2020) and molecular evidence (Poelstra

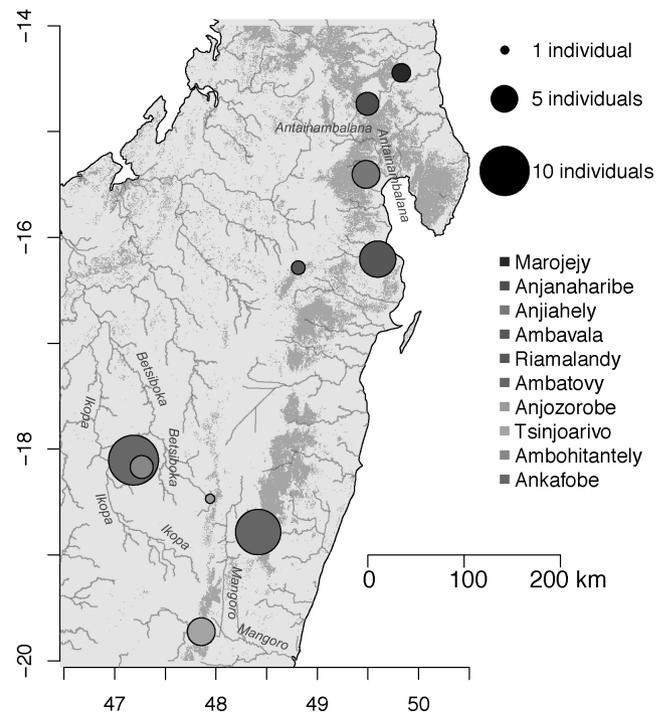
et al., 2021). Of additional paleoecological interest, *M. lehilahytsara* is found at higher altitudes than other mouse lemur species, perhaps due to its ability to enter a state of prolonged torpor (Blanco et al., 2017), thus reducing its metabolic demands during periods of extended low temperatures during the austral winter. To summarize, the species distribution and life history of *M. lehilahytsara* suggest that it has long been present in the Central Highlands and that demographic studies of these forest-dwelling primates may illuminate pre-human conditions in Madagascar. Population divergences can reflect forest fragmentation, and population size fluctuations through time will reflect pulses in forest distributions. Although loss of genetic diversity in endangered lemurs species can be attributed to human activities (Olivieri et al., 2008; Salmona et al., 2017; Teixeira, Montade, et al., 2021), a better informed reconstruction of prehuman conditions can reveal if some populations are especially vulnerable due to the effects of natural climate change (Quéméré et al., 2012; Salmona et al., 2017; Teixeira, Montade, et al., 2021).

Here, we sampled *M. lehilahytsara* populations from multiple sites in the Central Highlands as well as from eastern rainforests, which include formerly recognized *M. mittermeieri* populations in the northeast and sites that are further south but along the escarpment where rainforests transition to the Central Highlands and more open vegetation. Using phylogeographic approaches and coalescent methods to analyse a large restriction site associated DNA sequencing (RADseq) data set, we explored patterns of genetic connectivity among populations. We also estimated divergence times between populations and changes in effective population size ( $N_e$ ) through time. Our findings show that population divergences occurred well-before humans arrived in Madagascar. The initial isolation of the Central Highlands from eastern rainforests occurred even before the Last Interglacial period (LIG; c. 132–112 ka). Post-divergence gene flow among populations in the Central Highlands and eastern rainforests suggests that migration was possible despite hypothesized large-scale vegetative turnover from more closed to open ecosystems. We find that there have been two notable episodes of population size contraction; the first coinciding with the last glacial maximum (LGM; c. 26.5–19 ka) and the second coinciding with recent human colonization (c. 2 ka). These patterns suggest that a combination of paleoclimatic and anthropogenic effects have synergistically increased extinction risk in *M. lehilahytsara* – a phenomenon that undoubtedly applies to other forest-dwelling species.

## 2 | MATERIALS AND METHODS

### 2.1 | Study sites and sampling

*Microcebus lehilahytsara* individuals were captured at five field sites (Figure 1). Two locations, Ambohitantely and Ankafoabe, are located within the Central Highlands. Their vegetational components are characteristic of moist evergreen forests, although more open and drier. Both Ambohitantely and Ankafoabe are protected forested areas well-characterized by the extensive grasslands surrounding them. The three other locations, Anjozorobe, Ambatovy, and Tsinjoarivo,



**FIGURE 1** Sampling sites for *Microcebus lehilahytsara* populations. Forest cover in green is taken from Schübler, Mantilla-Contreras, et al. (2020), which reflects modern-day presence rather than historical connectivity. Water ways are shown in blue with relevant rivers named in italics. The x-axis is longitude, and the y-axis is latitude. [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/mec.16632)]

are mid-elevation rainforests at the southern end of the species range situated along the eastern escarpment (hereafter referred as southern [rainforest] populations; Figure S1). We complemented our sampling with an existing data set of five *M. lehilahytsara* populations at humid rainforest sites in northeastern Madagascar (Marojejy, Anjanaharibe-Sud, Anjahely, Ambavala, Riamalandy) reported in Poelstra et al. (2021) (hereafter referred as northern [rainforest] populations), where it was recommended that *M. mittermeieri* should be synonymized as *M. lehilahytsara*. Consequently, our sample set included 54 *M. lehilahytsara* individuals covering the entire known geographic range of the species. A detailed description of climate and topography of the sampling sites can be found in the Supporting Information Methods “Study Sites”. In addition, we included seven *M. tanosi* individuals from southeastern Madagascar and three *M. murinus* individuals from western Madagascar as outgroups. For a description of live-capture methods see Blanco et al. (2017). Collection information for all specimens is given in Table S1.

### 2.2 | RADseq library preparation and sequencing

We generated new RADseq data for 38 individuals while previously published sequences using the same methods were available for 26 individuals (Poelstra et al., 2021). RADseq library preparations generally followed Ali et al. (2016), using a single-digest protocol

with the SbfI restriction enzyme. A median insert size of 380 bp was selected for paired-end sequencing at the Duke Sequencing Core on an Illumina Hi-Seq 4000. Two libraries were prepared with the single-SbfI protocol described in Etter et al. (2011) and Genomic Resources Development Consortium et al. (2015). These two libraries were single-end sequenced on an Illumina HiSeq 2000 at the University of Oregon Core facility. Information on library preparation and sequencing for all new and previously published samples are given in Table S2.

### 2.3 | Data processing and genotyping

Raw RAD reads were demultiplexed with the `process_radtags` function in STACKS version 2.0b (Rochette et al., 2019), trimmed using Trimmomatic with the parameters Leading: 3, Trailing: 3, Slidingwindow: 4:15, and Minlen: 60 (Bolger et al., 2014), and aligned against the *Microcebus murinus* reference genome (Mmur 3.0; Larsen et al., 2017) using BWA version 7.0.17 with the BWA-MEM algorithm and default settings (Li, 2013). Trimmed reads of technical replicates, that were available for 16 samples (Tables S2), were combined prior to reference alignment to increase coverage. Subsequently, BAM files were filtered with SAMtools version 1.11 (Li et al., 2009) to retain only reads mapping to autosomal scaffolds with a minimum mapping quality of 20. Paired-end sequenced data was additionally filtered for proper pairing and deduplicated. SAMtools was also used to count the number of RAD loci sequenced for each individual and estimate locus coverage as the forward read depth at the respective SbfI cutting site.

Two genotyping approaches were employed to evaluate robustness of our results to genotyping errors. First, we used the reference-based approach of STACKS version 2.53 (Rochette et al., 2019), including only loci present in at least 75% of all individuals. Genotype calls were filtered using VCFtools version 0.1.17 (Danecek et al., 2011) following “FS6” recommendations of O’Leary et al. (2018) with modified thresholds (Methods “Genotype filtering”). A minor allele count filter of three was applied for analyses of population structure. Second, genotype likelihoods were estimated with the SAMtools model in ANGSD version 0.934 (Korneliussen et al., 2014), using filtering thresholds applied in closely-related mouse lemurs (Poelstra et al., 2021) and excluding *M. lehilahytsara* individuals that did not pass genotype filtering and outgroups (Supporting Information Methods “Genotype likelihood estimation with ANGSD”). Genotype likelihoods were used for analyses of population structure and to infer minor allele frequency (MAF) spectra for all populations and population pairs (i.e., joint MAF spectra) for demographic analyses, using realSFS implemented in the ANGSD framework.

### 2.4 | Phylogenetic inference

We estimated phylogenies using three different approaches with genotype calls, which allowed us to explore uncertainty in phylogeographic history due to model assumptions for each method:

1. Maximum likelihood (ML; Felsenstein, 1981) analysis with RAxML-NG (Kozlov et al., 2019) was carried out on the unpartitioned concatenated alignment of SNPs using the GTR+ $\Gamma$  model of molecular evolution (Tavaré, 1986; Yang, 1994) while correcting for ascertainment bias (Lewis, 2001). We conducted 20 unconstrained ML searches and performed 100 bootstrap replicates.
2. Topological estimation from invariants was performed with SVDquartets (Chifman & Kubatko, 2014) implemented through PAUP\* version 4.0a (build 168; Swofford, 2003), using a thinned data set with a minimum distance of 10,000 bp between SNPs to potentially avoid treating SNPs in linkage disequilibrium as independent. We evaluated all quartets with 100 standard bootstrap replicates and estimated phylogenies where each individual was a tip and where individuals were assigned to populations a priori based on sampling location.
3. SNAPP (Bryant et al., 2012), a Bayesian implementation of the multispecies coalescent (MSC; Rannala & Yang, 2003) model for SNP data, was run using BEAST2 version 2.4.8 (Bouckaert et al., 2014). Similar to SVDquartets, we used a thinned data set to avoid site independence violations. To reduce the computational burden, we selected only two individuals per sampling location based on mean sequencing depth (Tables S1 and S2). Four replicates were run to evaluate convergence. Mixing and effective sample sizes were analysed with Tracer version 1.7 (Rambaut et al., 2018). Details regarding priors and MCMC options are in the Supporting Information Methods “SNAPP priors and settings”.

To evaluate alternative topological hypotheses for ML and MSC methods, we used probabilistic topology hypothesis testing approaches. Specifically, we tested if the earliest split among *M. lehilahytsara* populations was between northern rainforest populations and southern rainforest plus Central Highland populations or between the Central Highlands and all rainforest populations. For ML, constrained optimizations of topologies and site likelihoods were performed with RAxML-NG, and support between the two competing topologies was evaluated with an approximately unbiased (AU; Shimodaira, 2004) test in IQ-TREE version 2.1.3 (Minh et al., 2020), using the GTR+ $\Gamma$  model and 10,000 RELL (Kishino et al., 1990) replicates. For SNAPP, we used stepping-stone integration (Xie et al., 2011) to obtain marginal likelihoods with 24 steps. Because differences in marginal likelihoods were large, we presented natural log Bayes factors (Kass & Raftery, 1995) as opposed to computing model probabilities.

### 2.5 | Population structure

Population structure was evaluated through clustering analyses, principal component analysis (PCA), and fixation index ( $F_{ST}$ ) statistics, leveraging both genotype calls and genotype likelihoods. Individual ancestries were inferred from genotype calls with STRUCTURE version 2.3.4 (Pritchard et al., 2000) and from genotype likelihoods

with ngsAdmix version 32 (Skotte et al., 2013). We conducted 20 independent runs for genotype calls and 10 runs for genotype likelihoods with the number of a priori clusters ( $K$ ) ranging from 1–10 and calculated  $\Delta K$  for estimating the most likely number of Hardy–Weinberg groups following Evanno et al. (2005). PCA was conducted on genotype calls using the R package adegenet version 2.1.3 (Jombart & Ahmed, 2011) and on genotype likelihoods using PCAngsd version 1.01 (Meisner & Albrechtsen, 2018). Pairwise  $F_{ST}$  between populations were calculated from genotype calls as described by Weir and Cockerham (1984) using the R package hierfstat version 0.5-7 (Goudet, 2005) and from genotype likelihood-based joint MAF spectra using realSFS with weighted  $F_{ST}$  calculations (Reynolds et al., 1983). The populations Riamalandy and Anjzorobe were excluded for  $F_{ST}$  estimation because only two and one individuals were available for them, respectively. Mantel tests were conducted at the population level using the R package vegan version 2.5-7 (Oksanen et al., 2020) to test the presence of isolation-by-distance using 9,999 permutations. We tested three alternative sets of populations: (1) northern rainforests only, (2) southern rainforests plus Central Highlands, and (3) all populations. Physical distances between populations were calculated with the Geographic Distance Matrix Generator version 1.2.3 (Ersts, 2021). Pairwise differences between individuals were used to perform an AMOVA analysis (Excoffier et al., 1992) with the R package poppr version 2.8.7 (Kamvar et al., 2014), grouping populations into the northern rainforests, southern rainforests, and Central Highlands to evaluate sources of variation.

## 2.6 | Demographic modelling

Genotype likelihoods can alleviate known biases of RADseq data and improve demographic inference from allele frequencies (Heller et al., 2021; Pedersen et al., 2018; Warmuth & Ellegren, 2019) because they include information about the uncertainty associated with genotype calls. Especially for low coverage data it may be important to account for this uncertainty, as stochastic under- and overrepresentation of alleles can impair genotype calling (Nielsen et al., 2011). Therefore, joint MAF spectra estimated from genotype likelihoods were used to test models of population splitting, population size change, and gene flow with fastsimcoal2 version 2.6.0.3 (Excoffier et al., 2013). For each model, we used 100 independent approximations with 200,000 simulations and 40 cycles of the expectation maximization algorithm. The best likelihood for each model was used to calculate its Akaike information criterion (AIC; Akaike, 1974) score. We used the AIC scores to calculate model probabilities such that model  $w$  has probabil-

ity  $\frac{\exp\{AIC_w - AIC_{min}\}}{\sum_i^n \exp\{AIC_i - AIC_{min}\}}$ . Joint MAF spectra models were calibrated to absolute time using a per-generation mutation rate of  $1.52 \times 10^{-8}$  based on a recent de novo mutation study with a *M. murinus* pedigree (Campbell et al., 2021) and a generation time of 3.5 years

based on parentage and survival estimations from wild populations of *M. murinus* (Radespiel et al., 2019) and *M. rufus* (Zohdy et al., 2014). We consider both studies here by averaging the two (i.e., taking the midpoint) since generation time differences between species may reflect present-day resource availability rather than inherited life history traits.

Because joint modelling of all populations at once would lead to an untenable computational burden for model selection, populations were split into three sets of models that explored demographic histories of (1) Ambavala from the northern rainforests versus previously recognized *M. mittermeieri* (Anjanaharibe and Anjiahely), (2) Ambavala from the northern rainforests versus Ambatovy and Tsinjoarivo from the southern rainforests, and (3) Ambatovy and Tsinjoarivo from the southern rainforests versus Ankafobe and Ambohitantely in the Central Highlands. Complexity was added to models in a hierarchical fashion to constrain the total number of model comparisons (Supporting Information Methods “Demographic model selection”). A final set of models including all seven populations evaluated among subsets was run for parameter estimation. We used 200 nonparametric block-bootstraps (Supporting Information Methods “Block-bootstrapping MAF spectra”) to estimate percentile-based 95% confidence intervals for the maximum pseudo-likelihood estimates.

## 2.7 | Characterizing population size changes through time

Population size change through time was evaluated with stairway plots (Liu & Fu, 2015). Similar to our other demographic analyses, we used genotype likelihood-based MAF spectra since there was no suitable outgroup to determine the ancestral state of each allele (Teixeira, Montade, et al., 2021). Because not all populations had appropriate sample sizes for stairway plot analyses, we focused on four representative populations: Ankafobe for the Central Highlands, Ambatovy for the southern rainforests, Ambavala for the northern rainforests, and a metapopulation of all formerly named *M. mittermeieri* individuals from the northern limit of the species distribution. For each population, we also generated 199 bootstrap replicates to obtain 95% confidence intervals, using the script provided by stairway plot version 2.0 (Liu & Fu, 2020). Analyses were rescaled from generations to absolute time given a per-generation mutation rate of  $1.52 \times 10^{-8}$  and generation time of 3.5 years.

Because stairway plots are sensitive to the number of coalescent events, which is dependent on the number of individuals and SNPs in the data set (Foote et al., 2016; Lapierre et al., 2017; Patton et al., 2019; Teixeira, Salmons, et al., 2021), analyses were initially performed considering the entire population data sets ( $n$  ranging between 7 and 12) and repeated with an equal sample size ( $n = 7$ ) to avoid differences in  $N_e$  among populations related to sample size. For the subsampled analyses, individuals were randomly selected (Table S1).

## 2.8 | Identifying potential migration corridors along rivers

We used ArcMap of ArcGis Desktop version 10.6.1 (ESRI Inc.) to visualize potential migration corridors along rivers by creating 10 m resolution false-colour composites (infrared, red green, bands 8-4-3) from tiles T38KQE, T38KQF, T38KRD and T38KRE of Sentinel-2 satellite images (Multi-Spectral Instrument, Bottom-of-Atmosphere reflectance [S2MSI-2A]) taken on 23 August 2021 and acquired from the European Space Agency sci-hub (<https://scihub.copernicus.eu/>).

## 3 | RESULTS

### 3.1 | RADseq data statistics

On average per individual, 8,181,656 raw reads were sequenced, of which 35.6% passed filtering and aligned to autosomal scaffolds (Table S2). Genotyping with STACKS and subsequent filtering resulted in a data set with 217,273 sites across 59 individuals (including 9 outgroup individuals) with an average 15 $\times$  coverage per individual. After the minor allele count filter, 111,104 sites were retained for downstream analyses. Genotype likelihoods estimated with ANGSD were available for 514,309 sites after filtering. Genotyping statistics can be found in Table S3 and percentage of missing data per individual is given in Table S4.

### 3.2 | Phylogenetic inference

All phylogenetic analyses recovered a well-supported split between northern rainforest and southern rainforest plus Central Highland *M. lehilahytsara* populations (Figure 2; Figures S2 and S3). There were minor incongruences between methods relating to the topology of the latter clade: MSC analyses (SNAPP and SVDquartets) found a well-supported split between Central Highland (Ambohitantely and Ankafobe) and southern rainforest (Ambatovy, Anjozorobe, and Tsinjoarivo) localities (Figure 2c; Figures S2 and S3) while the ML analysis (RaxML) found a well-supported clade comprising Anjozorobe and Central Highland populations (Figure 2a). ML analyses recovered sampling locations as clades, except for Ambatovy, which was paraphyletic with weak support, such that Tsinjoarivo was nested within (Figure 2a). In contrast, SVDquartets analyses of all individuals found Ambatovy and Tsinjoarivo to form two well-supported clades (Figure S2). Furthermore, MSC-based analyses were not in complete agreement: SNAPP recovered Riamalandy and Ambavala as sister (Figure 2c) while SVDquartets recovered Riamalandy and Ambavala as a grade, albeit, with poor (Figure S2) or weak support (Figure S3).

Because the split order between populations is of consequence for downstream demographic modelling, we also explored the strength of evidence in favour of a north-versus-south/highlands

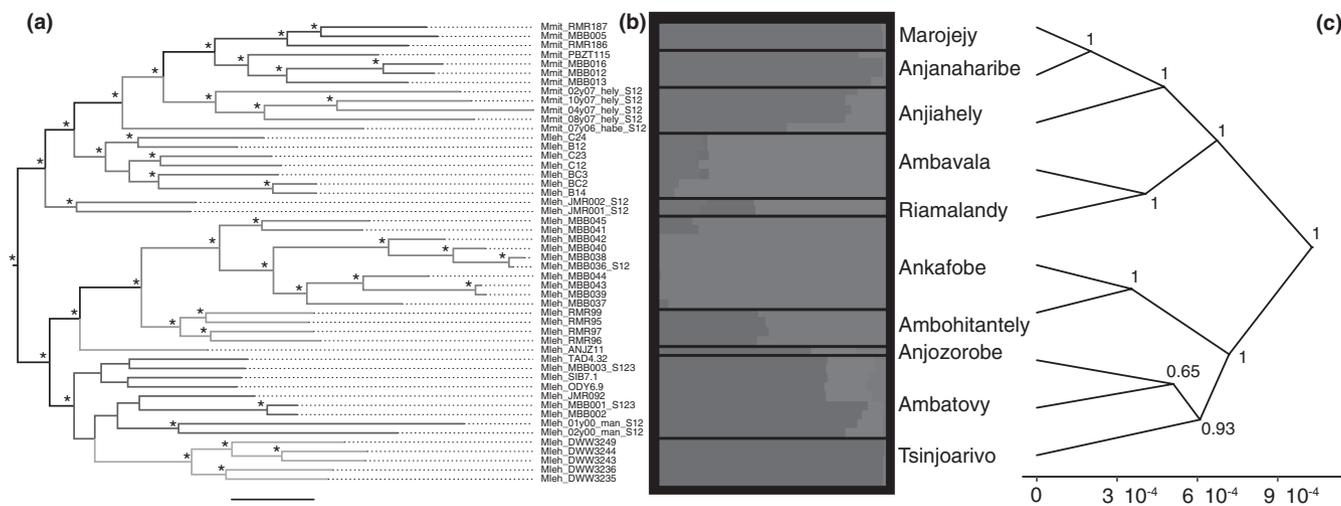
hypothesis relative to a highlands-versus-humid rainforests hypothesis. ML-based AU tests rejected the alternative scenario of a highlands-versus-humid rainforests basal split ( $p$ -value = .0179). Similarly, marginal likelihoods from SNAPP decisively favoured the north-versus-south/highlands model (marginal ML = -82860.30) over the highlands-versus-humid rainforests model (marginal ML = -82889.96) with a Bayes factor of -59.32.

### 3.3 | Delimiting population structure

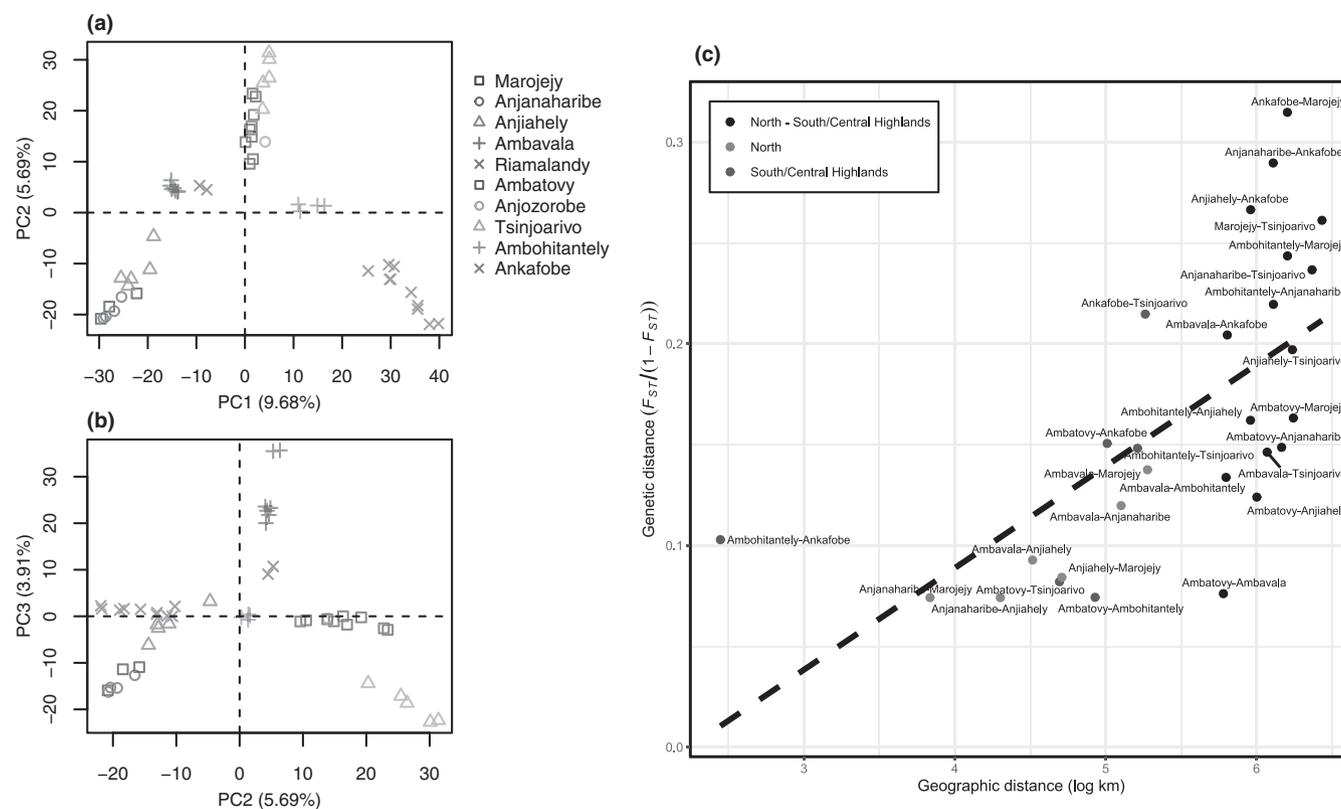
Population structure mirrored findings from phylogenetic analyses and revealed admixture between *M. lehilahytsara* populations in the Central Highlands and southern rainforests, and between the southern and northern rainforest populations (Figure 2b). An optimal  $K$  of four was selected by STRUCTURE analyses of hard-filtered genotypes (Table S5), which yielded very similar results to the optimal  $K$  of three for genotype likelihoods from ngsAdmix (Figure S4; Table S6). Notably, Ambohitantely had near 50:50 assignment to clusters from the Central Highlands and rainforest populations, represented by Ankafobe and Ambatovy, respectively (Figure 2b and Figure S5). Populations previously identified as *M. mittermeieri* largely formed their own cluster, although some admixture between Anjahely (south of the Antainambalana river) and Ambavala and between Anjahely and Anjanaharibe (north of the Antainambalana river) was prevalent at higher  $K$ . Increasing  $K$  did not always recover the expected geographic groupings as separate clusters (Figures S5-S7).

PCA supported the STRUCTURE and ngsAdmix clustering results. PC1 generally separated northern rainforest, southern rainforest, and Central Highland populations. PC2 further separated southern rainforest populations from the northern rainforests and Central Highlands (Figure 3a). Ambohitantely took an intermediate position between Ankafobe and Ambatovy while Ambavala and Riamalandy fell between other northern rainforest populations and Ambatovy. PC3 further separated Tsinjoarivo from Ambatovy, and showed clustering among individual northern rainforest populations (Figure 3b). Analyses with genotype likelihoods found similar patterns (Figure S8). Significant geographic structure was detected between populations with AMOVA analyses (Table S7) although geographic sampling location explained only about 8% of genetic variation.

Pairwise  $F_{ST}$  values (Table S8) regressed against Euclidean distances (Table S9) indicated the presence of isolation-by-distance when comparing all populations with a Mantel test (Pearson's  $r = .78$ ,  $p = 7 \times 10^{-4}$ ; Table S10) (Figure 3c). This relationship was weaker when only considering  $F_{ST}$  estimates between northern rainforests and southern/Central Highland populations (Pearson's  $r = .49$ ,  $p = .15$ ; Table S9). Several data points stood out as inconsistent with a simple IBD pattern. For example, genetic distance between Ankafobe and Ambohitantely was higher than between Ambatovy and Ambohitantely despite their phylogenetic placement as sisters and being only 12 km apart as opposed to 138 km. Similarly, the genetic distance between Ambatovy and Ambavala was much lower



**FIGURE 2** Well-supported phylogenetic lineages of *Microcebus lehilahytsara* reflect genetic structure. (a) The ML phylogeny of all individuals indicates well-supported nodes (bootstrap support  $\geq 70\%$ ) with an asterisk (\*), while poorly supported nodes (bootstrap support  $< 70\%$ ) are blank. Branch colours correspond to population colours in Figure 1. Branch lengths are expected substitutions per site. (b) A STRUCTURE analysis of all individuals reflects supported clades that correspond to geography at an optimal  $K$  of four. (c) A full likelihood MSC analysis with SNAPP further supports the ML topology and major splits with strong posterior probabilities, with the exception of Anjozorobe, which is represented by a single individual. Branch lengths are expected substitutions per site. [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]



**FIGURE 3** Principal component analysis (PCA) of genotype data and pairwise  $F_{ST}$ . (a) The first and second axes generally separate northern, southern, and Central Highland populations. (b) The third axis further separates northern populations and Tsinjoarivo from Ambatovy. (c) Pairwise  $F_{ST}$  values between populations reveal evidence for IBD, but several points deviate from the trend that may be explained by gene flow. The dashed line represents the fitted linear regression. [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]

than expected from the regression model, which was congruent with evidence of admixture from STRUCTURE, ngsAdmix, and PCA results. Previously mentioned patterns were not always consistent with pairwise  $F_{ST}$  estimates from genotype likelihoods (Figure S9), but results of Mantel tests (Table S9) similarly imply distance is not the sole explanation of observed genetic differentiation.

### 3.4 | Demographic modelling

Plausible models based on phylogenetic and population structure analyses were explored to understand historical and contemporary gene flow among northern populations (Ambavala, Anjahely, and Anjanaharibe; Figure S10), between north and south (Ambavala vs. Ambatovy, and Tsinjoarivo; Figure S11), and between the Central Highlands and southern rainforests (Ankafobe and Ambohitantely vs. Ambatovy and Tsinjoarivo; Figures S12 and S13). The best models for each data set (Tables S11–S13) revealed that divergences between sister populations probably occurred sometime before human colonization of the Central Highlands and eastern Madagascar but after the LGM, even though confidence intervals were too wide to reject post-human arrival divergences (Figure 4). The models for northern populations (Figure 4a) and for the comparison of northern and southern populations (Figure 4b) detected a two-order of magnitude decline in  $N_e$ , except for Ambatovy, which probably occurred after human colonization, but again, confidence intervals did not rule out early Holocene or late Pleistocene declines. No such crash was detected for the analysis of Central Highland and southern populations (Figure 4c). Rather, there was evidence for both historical and recent gene flow between the Central Highlands and Ambatovy. However, confidence intervals on migration rate parameters were typically wide, and there was no discernable pattern among migration rates through time to determine whether gene flow has actually decreased towards the present between the Central Highlands and Ambatovy (Figure 4c). Model selection results suggested that only historical gene flow was detectable between the north and south, which stopped shortly after the divergence of Ambatovy and Tsinjoarivo (Figure 4b), and that historical gene flow occurred between Ambavala and the common ancestor of Anjanaharibe and Anjahely (Figure 4a).

Because confidence intervals were typically wide causing ambiguous interpretations and root divergence times were ultimately in conflict with the population tree (Figure 2c) split order (Tables S14–S16), we aimed to improve parameter estimation by jointly modelling all seven populations at once. This required some additional model testing to determine the population splitting order with gene flow (Figures S14 and S15; Table S17), and gene flow patterns observed in smaller models were assumed correct. Ancient population structure (i.e., the northern, southern, and Central Highland populations) formed at least before the LGM and probably prior to LIG (Figure 5). Subsequent population divergences occurred near the LGM, or at least the LGM was contained within confidence intervals while plausible human effects were excluded. The end of gene flow between northern and southern populations (i.e., Ambavala and Ambatovy) as

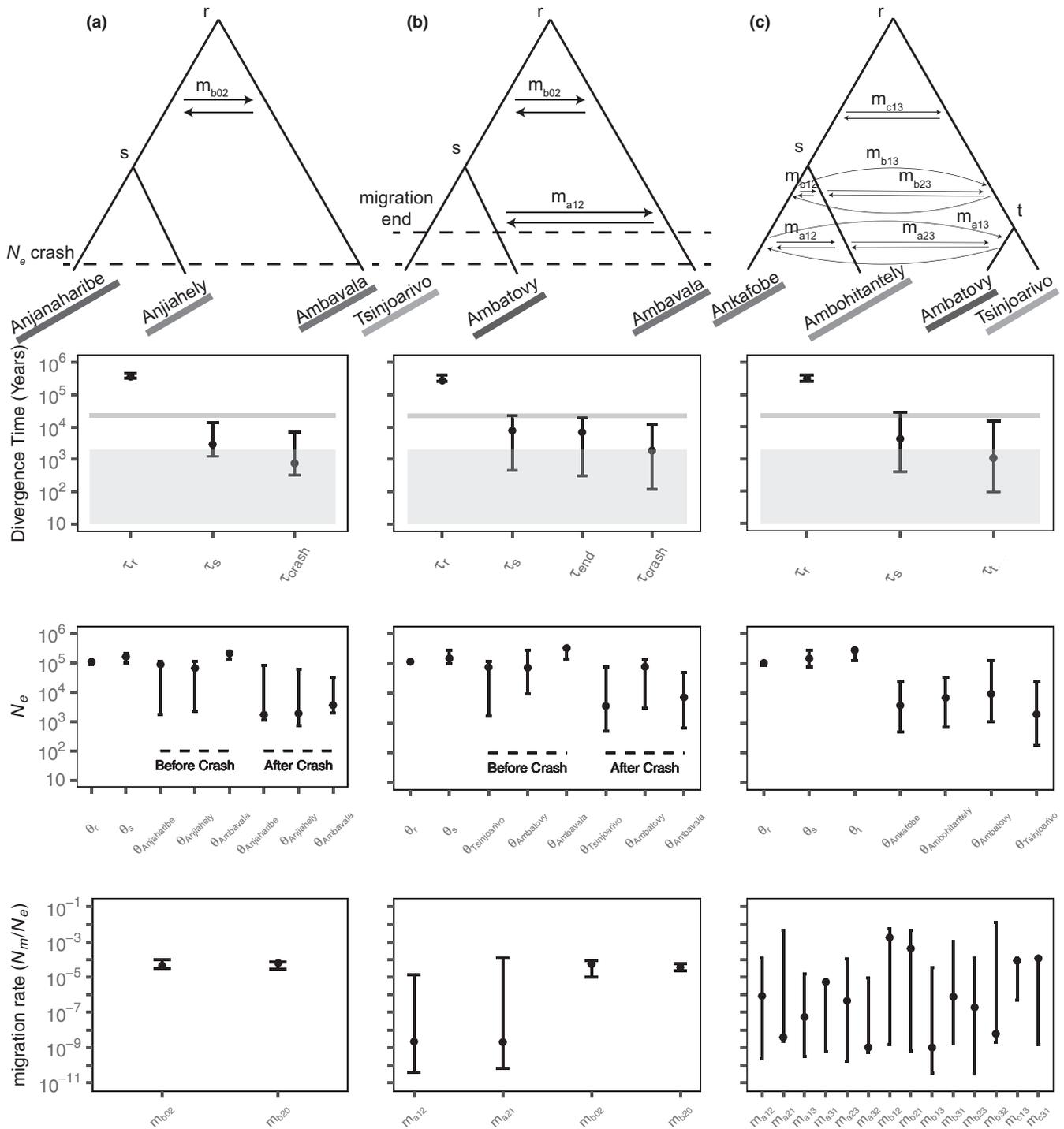
well as an  $N_e$  crash experienced by almost all populations occurred closer to the timing of human colonization, but potentially in the late Pleistocene after the LGM. Nearly a 10-fold  $N_e$  decrease was estimated for all populations from the modelled recent  $N_e$  change with the exception of Ambatovy (Figure 5; Table S18). There was no remarkable pattern among the migration rate parameters except that confidence intervals imply a high degree of uncertainty in migration rate point estimates. For a few parameters, the maximum pseudo-likelihood estimates fall outside of their confidence intervals, but not for divergence times or more recent  $N_e$  estimates.

### 3.5 | Characterizing population size changes through time

Similar demographic trends were recovered for the representative populations when using the same number of individuals per population (Figure 6). Assuming that undetected population structure is negligible, stairway plot inferred a period of constant  $N_e$  that predated the LIG and lasted until the LGM, with  $N_e$  lowest for the northern metapopulation formerly described as *M. mittermeieri* and highest for Ambatovy. After the LGM, there was a continuous decline in  $N_e$  until present for all forest sites except for the former *M. mittermeieri* metapopulation, where declines were delayed until the mid-Holocene at approximately 7.5 ka. All populations show a second decline during or just before a hypothesized period of human colonization and anthropogenic landscape changes (c. 2 ka; Figure 6). Despite the large confidence intervals observed for the analyses considering seven individuals, our results support the presence of both pre- and post-human colonization population declines (Figure S16). The stairway plot analyses including more than seven individuals where possible confirmed sensitivity of  $N_e$  estimates to sample size. Nevertheless, the same general patterns were observed compared to analyses with seven individuals (Figure S17), while recovering more fine-scale changes. Because of site filters for missing data, analyses with more than seven individuals ultimately had less data available for MAF spectra estimation than analyses subsampled to seven individuals (Table S19).

### 3.6 | Detecting potential migration corridors between the Central Highlands and southern rainforest populations

The Central Highlands of Madagascar are traversed by westward flowing rivers (Figure 7k), which were characterized by riparian forest or shrub vegetation before widespread transformation into agricultural fields (Kull, 2012). The Betsiboka river with its tributaries connects the humid rainforest population of *M. lehilahytsara* at Anjozorobe with the now isolated highland populations in Ambohitantely and Ankafobe. Remnants of riparian forest are still present in a few places, but most forests are converted into rice paddies today (Kull, 2012). The exchange of individuals using historically forested corridors along the Betsiboka river would also

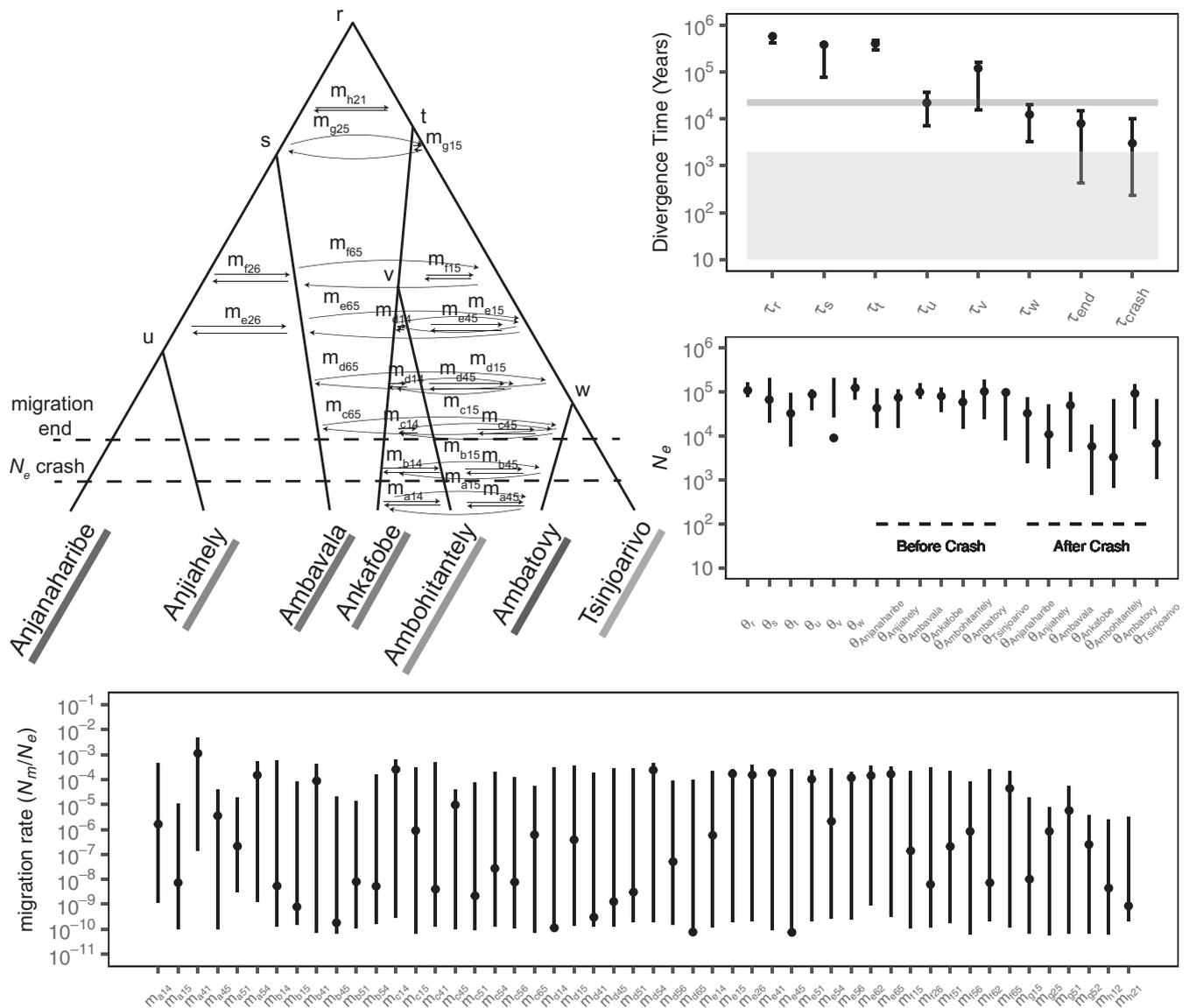


**FIGURE 4** Demographic models reveal the history of divergence, gene flow, and population size among studied populations. Selected SFS models for evaluating population divergence, population size change, and gene flow (a) among northern, (b) between northern and southern, and (c) between the Central Highland and southern rainforest populations. All parameter estimates are shown on a  $\log_{10}$  scale to reflect changes in orders of magnitude. Points are maximum pseudo-likelihood estimates and error bars are percentile-based 95% confidence intervals. Colours for divergence times are the LGM (blue) and the time since human colonization (yellow). [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/mec.16632)]

explain observed gene flow between southern rainforest populations such as Ambatovy and the Central Highlands.

However, the southern populations of *M. lehilahytsara* are separated by one large and several smaller rivers (Figure 7k). While it is likely that populations at Ambatovy and Anjozorobe (c. 60 km apart) could have been connected across the headwaters of the Mangoro

river (Figure 7j), Anjozorobe and Tsinjoarivo are already separated by c. 140 km of rainforest traversed by several smaller eastward flowing rivers with their sources in the Central Highlands. Finally, Ambatovy and Tsinjoarivo are separated by the very wide Mangoro river which is, apart from its source region, probably too large to be crossed by mouse lemurs (Figure 7h–j).



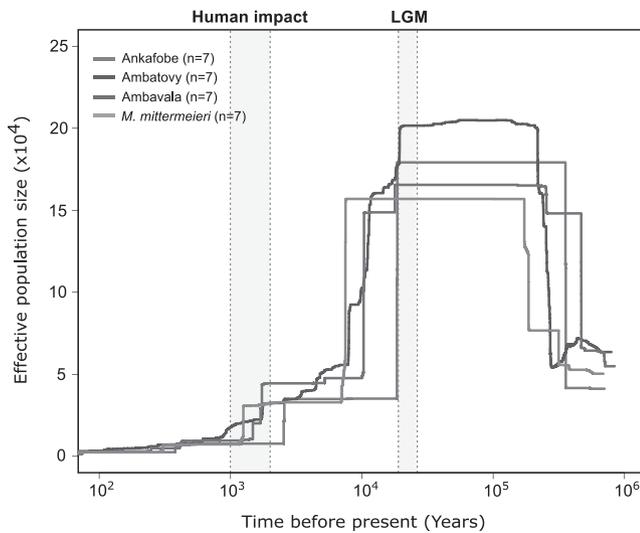
**FIGURE 5** Parameter estimates for the seven-olupation model inferred with fastsimcoal2. Each node is labelled *r* through *w*, which has a divergence time and  $N_e$  parameter associated with it.  $N_e$  parameters are indicated by  $\theta$  to avoid the extra subscript. Points are maximum pseudo-likelihood estimates and error bars are 95% confidence intervals. For the divergence time plot, the yellow colour indicates post-human colonization of Madagascar, and the blue colour indicates the timing of the LGM. All y-axes were transformed to a  $\log_{10}$  scale to better visualize order-of-magnitude changes. [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/mec.16632)]

## 4 | DISCUSSION

### 4.1 | Population structure of *Microcebus lehilahytsara* predates human colonization of Madagascar

Our results indicated that *M. lehilahytsara* Central Highland populations separated from rainforest populations long before human colonization of these regions (c. 2 ka; e.g., Douglass et al., 2019) and predate even the oldest potential human presence in Madagascar (c. 10 ka; Hansford et al., 2018). Our demographic models indicated that this divergence and the distinct genetic clusters (Figure 2b; Figure 3a) could have originated as far back as 500 ka, which may be associated with older Pleistocene interglacial cycles and the fragmentation of a

previously more continuous habitat (Figure 5). Divergences among populations within the northern, southern, and Central Highland clusters likely originated sometime between the LGM and mid-Holocene (Figures 4 and 5). Charcoal records (Burney, 1987a, 1987b) and stratigraphic pollen occurrence data (Burns et al., 2016; Gasse & Van Campo, 1998; Virah-Sawmy et al., 2010) have shown a period of increased drought and fire from about 6 ka to the present. The LGM also induced cool and dry conditions in Madagascar (Clark et al., 2009; Scholz et al., 2011; Teixeira, Montade, et al., 2021), which were characterized by more open habitats in the Central Highlands (Burney et al., 2004; Samonds et al., 2019), even though it is debatable what the vegetational components of that open habitat were (e.g., Joseph et al., 2021). During this dry period, *M. lehilahytsara*, which is currently associated with humid environments, likely



**FIGURE 6** Population size change of *Microcebus lehilahytsara* through time estimated with stairway plots. The same sample size ( $n = 7$ ) was used for all populations. The metapopulation of Anjahely, Anjanaharibe and Marojejy northern rainforest sites is labelled *M. mittermeieri* to ease legend formatting although these individuals are now recognized as *M. lehilahytsara*. Absolute time calibrations were a generation time of 3.5 years and a per-generation mutation rate of  $1.52 \times 10^{-8}$ . The vertical bars identify the LGM between 19 and 26.5 ka, and the start of a significant human impact in Madagascar between 1 and 2 ka. [Colour figure can be viewed at [wileyonlinelibrary.com](https://onlinelibrary.wiley.com/doi/10.1111/mec.16632)]

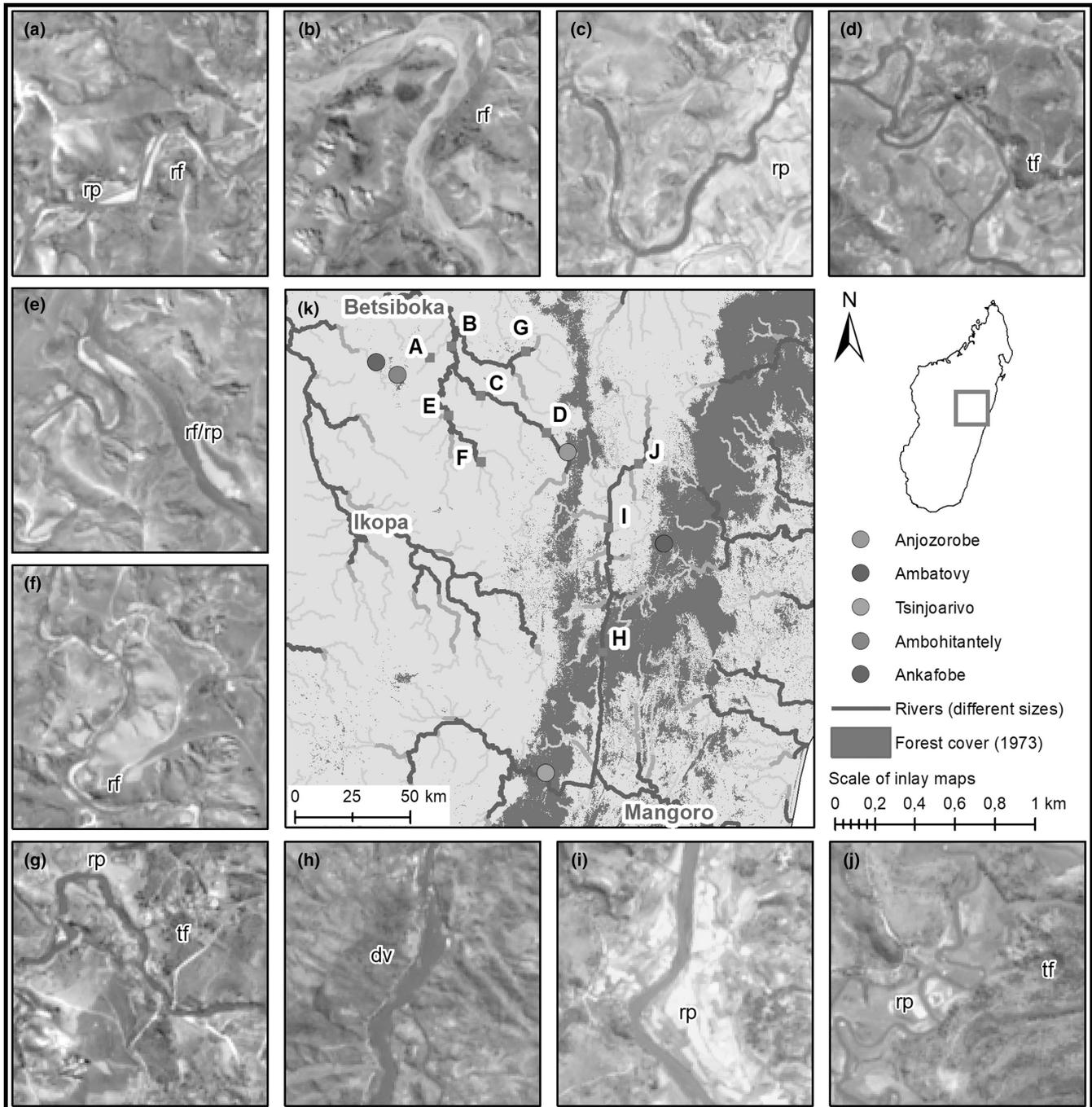
retreated to watersheds buffered from aridification (Muldoon et al., 2012; Wilmé et al., 2006). This would explain the near simultaneous population separations in the north, south, and Central Highland lineages. Although the ML estimate and confidence interval show uncertainty in the divergence time between Ankafobe and Ambohitantely, this may reflect the complex patterns of post-divergence gene flow between the Central Highlands and southern rainforest populations.

Rivers are known to function as barriers to gene flow in mouse lemurs (Olivieri et al., 2007) and other lemurs (Quéméré et al., 2010), and our data suggest that they are more significant in impeding mouse lemur migration than open grasslands. For example, Tsinjoarivo is isolated by the Mangoro river despite recent connectivity of closed-canopy forest habitats with Ambatovy (Figures 1 and 7). Analyses that support the Mangoro as a gene flow barrier include findings that individuals from Tsinjoarivo formed a well supported clade in the phylogenetic analyses (Figures 2 and S2), a distinct cluster in ngsAdmix analyses when increasing  $K$  from four to five (Figure S6) and were recognizable in PCAs along PC axis 3 with both hard-filtered genotypes and genotype likelihoods (Figure 3; Figure S8). Similarly prominent was the divergence of Anjahely from Anjanaharibe and Marojejy in the north. Anjahely lies south of the Antainambalana river system while Anjanaharibe and Marojejy are to the north (Figure 1). There was no evidence of post-divergence gene flow between Anjahely and Anjanaharibe (Figure 4), and although this region is characterized by heavy deforestation since the 1990s (Schüßler, Mantilla-Contreras, et al., 2020), their divergence time was

estimated to occur near the LGM and at least before the Holocene (Figure 5; Table S17). This divergence time (between Anjahely and Anjanaharibe) suggests that barriers to gene flow existed well before recent deforestation, even if the generation time used for calibrating analyses was overestimated by a factor of three. The phylogenetic and population genetic evidence combined with our best demographic models thus favour a scenario of prehuman climate-driven population divergence through the Pleistocene with rivers preventing gene flow even when precipitation increased following dry periods. There is a limited role for IBD explaining population structure in *M. lehilahytsara* (Figure 3; Figure S9; Table S9) which is also indicated by some recent analyses of mouse lemurs including *M. lehilahytsara* from northeastern Madagascar (Poelstra et al., 2021). However, distance alone has not prevented gene flow across the seemingly open habitats of the Central Highlands through time.

## 4.2 | The Central Highland mosaic of grassland and wooded savanna was permeable

Three lines of evidence support the hypothesis that *M. lehilahytsara* had a large ancestral distribution that covered the Central Highlands and the eastern rainforests. Within this large geographic area, population substructure was formed by paleoclimatic oscillations but with the maintenance of some level of post-divergence gene flow between Central Highland and southern rainforest populations. First, based on our coalescent estimates, the initial divergence between the Central Highland and southern rainforest populations occurred near-simultaneously with the divergence between their common ancestor and the northern rainforest populations (Figure 5). Confidence intervals reflect uncertainty in the timing of these divergences, but a shared demographic history among populations is also evident in the stairway plot analyses (Figure 6). Second, were the Central Highland populations simply a small founder population that dispersed from the eastern escarpment rainforests much later, we would expect signatures of a bottleneck and recovery to be observed rather than a similar trend of large ancestral population sizes accompanied with Pleistocene declines. The shared demographic trends are robust to sample size (Figure S17) and support the ancient origins of *M. lehilahytsara* in the Central Highlands, even though stairway plots are less accurate at reconstructing ancient histories (e.g., thousands of generations) compared to recent ones (e.g., hundreds of generations; Patton et al., 2019). There are large disparities in  $N_e$  estimates between stairway plot and fastsimcoal2 analyses, even when considering a recent anthropogenic  $N_e$  crash across all populations (Figure 6; Table S17). Absolute  $N_e$  values should be interpreted with caution; however, the patterns of  $N_e$  change through time inferred from the different analyses are generally consistent when considering confidence interval widths. Third, the strong preference for continuous gene flow between Ankafobe, Ambohitantely, and Ambatovy (Figure 4; Figures S12 and S13; Table S12) suggests movement across the Central Highland habitat through time, even compared to models that allowed for some post-divergence gene flow



**FIGURE 7** Potential riparian corridors along the Betsiboka and Mangoro Rivers. Riparian corridors potentially link rainforest and Central Highland populations of *Microcebus lehilahytsara* (k). The inlays (a–j) illustrate the presence of riparian forests (rf), riparian areas that were converted into rice paddies (rp) but were forested before (Kull, 2012), transitional forests (tf) between rainforest and riparian highland vegetation and degraded vegetation (dv) along the two rivers. Inlays are produced from Sentinel-2 false colour composites. Rivers were derived from Schübler, Mantilla-Contreras, et al. (2020) and forest cover was taken from Vieilledent et al. (2018). The images show in blue-turquoise colour the river courses, in light green-beige grassland vegetation and bare soil, in pale-slightly red colour with regular shapes rice paddies, in light red shrubland, thin riparian vegetation narrower than the resolution of 10 m, and in rich red transitional forest vegetation or riparian forests. [Colour figure can be viewed at [wileyonlinelibrary.com](http://wileyonlinelibrary.com)]

but not until the present. Previous research has also suggested that mouse lemurs were capable of moving across the Central Highlands during the Pleistocene based on phylogeographic inference (Yoder et al., 2016), but this would presumably not be possible after human-mediated fire-driven expansion of grasslands (Burney et al., 2004;

Gasse & Van Campo, 1998). Our gene flow model is supported by the identification of riparian forest corridors (Figure 7) that would have enabled movements between the Central Highlands and the eastern escarpment rainforests. Such corridors may also have been key to previous mouse lemur dispersal events across a grassland and

woodland mosaic. It is unlikely that substantial migration between the studied Central Highland and southern rainforest populations persists today as the rapid deforestation of Madagascar's eastern rainforests in recent years has resulted and will further result in the fragmentation of forest corridors (Schüßler, Mantilla-Contreras, et al., 2020; Vieilledent et al., 2018).

Some caution is needed in the interpretation of our demographic models, as there is little discernable direction in gene flow, and confidence intervals for migration rate parameters typically span several orders of magnitude. This shows some limitations of our analyses, as we are obtaining estimates under a parameter-rich model, where there can be identifiability problems given the many combinations of divergence times,  $N_e$ , and migration rates that can explain the observed MAF spectra. However, several other observations are consistent with our model such as the higher-than-expected genetic distance between Ankafoke and Ambohitantely (Figure 3; Figure S9), model-based clustering analyses (Figure 2; Figures S5 and S6), and the intermediate position of Ambohitantely between Ankafoke and Ambatovy in PCAs (Figure 3; Figure S8). Additional sampling of individuals may be necessary to potentially reduce confidence interval widths. In addition, having population-level representation of Anjozorobe would resolve the site's current phylogenetic ambiguity (Figure 2; Figures S2 and S3) while filling a vital sampling gap along the proposed river corridors (Figure 7).

### 4.3 | Making conservation biology predictions from models of molecular evolution

Here, we provided a range of present-day  $N_e$  estimates that are sensitive to model choice, though the direction of change in  $N_e$  through time is not. Every analysis shows that *M. lehilahytsara* populations sizes have declined. Our SFS model selection and parameter estimation have been rigorous, and population substructure was accounted for in stairway plot analyses, since historical admixture or structure will manifest as  $N_e$  changes in single-population methods (e.g., Chikhi et al., 2010; Mazet et al., 2016). All evidence points to drastic population declines following the LGM as well as more recent declines coincident with human activities. These findings are consistent with previous studies of an endemic olive tree (Salmona et al., 2020), a mouse lemur (Teixeira, Montade, et al., 2021), and two sifaka species (Quéméré et al., 2012; Salmona et al., 2017), where population structure, as a proxy for forest fragmentation, and population declines were attributed to both prehuman climatic and later human effects in northern Madagascar. The most recent IUCN evaluation of *M. lehilahytsara* considers the species to be vulnerable (Dolch et al., 2020) while other mouse lemur species in the eastern rainforest are considered endangered or critically endangered (Andriaholinirina et al., 2014a, 2014b; Borgerson et al., 2020). We surmise that though *M. lehilahytsara* appears to be more robust to climate-driven and human-mediated disturbances than other species, this is more likely a consequence of its large ancestral geographic range combined with its physiological ability to inhabit both the lowlands and

higher altitudes. It was previously regarded as a highland specialist (Radespiel et al., 2012) but is now also recognized in lowland areas in the northeast after synonymizing *M. mittermeieri* (Schüßler, Mantilla-Contreras, et al., 2020). *M. lehilahytsara* has been documented to enter a state of prolonged torpor (Blanco et al., 2017) which may either be an adaptive trait that has facilitated its increased geographic range compared to most other mouse lemur species, or a plastic one that is only evident at higher altitudes. The importance of torpor for sustaining population sizes despite limited resources remains unclear. Regardless, the downward trends in  $N_e$  across the Central Highlands and the studied rainforest populations imply a negative consequence of continued habitat loss and fragmentation.

### 4.4 | The importance of interpreting biodiversity through evolution

The processes that shaped current *M. lehilahytsara* population structure must have occurred before human arrival and mid-Holocene droughts. Although charcoal records (Burney, 1987a, 1987b), stratigraphic pollen occurrence (Burns et al., 2016; Gasse & Van Campo, 1998), and megafauna subfossil extinctions (Hansford et al., 2018) all point to human-mediated large-scale ecological changes in the Central Highlands (Virah-Sawmy et al., 2010), our analyses suggest that *M. lehilahytsara* population divergences coincided with Pleistocene climate fluctuations both before and near the LGM. Our results, especially when combined with the narrative from other analyses of lemur populations (Quéméré et al., 2012; Salmona et al., 2017; Teixeira, Montade, et al., 2021) and endemic olive trees (Salmona et al., 2020), reveal a timeline for vegetational change in Madagascar from a population genetic perspective. Analyses of multiple species and localities agree with ecological (Bond et al., 2008) and evolutionary (Hackel et al., 2018; Vorontsova et al., 2016) studies of grasses that demonstrate Madagascar's Central Highland grasslands are of ancient origin, and further, that the Central Highlands have long persisted as a mix of grassland and woodland. Although fires due to human activity are prominent across the Central Highlands (Burney, 1987a; Burns et al., 2016; Gasse & Van Campo, 1998; Samonds et al., 2019; Virah-Sawmy et al., 2010), leading to devastating forest loss in Madagascar (Vieilledent et al., 2018), the effects of ongoing habitat fragmentation are occurring on a background of habitat mosaicism that is probably associated with Pleistocene climate change.

### AUTHOR CONTRIBUTIONS

George P. Tiley and Anne D. Yoder conceived the study. Marina B. Blanco, Edward E. Louis, José M. Ralison, Blanchard Randrianambinina, and Rodin M. Rasoloarison performed the fieldwork. Amanda R. Stahlke and Paul A. Hohenlohe constructed RADseq libraries. George P. Tiley, Tobias van Elst, Helena Texeira, and Dominik Schüßler performed analyses. George P. Tiley, Tobias van Elst, and Anne D. Yoder wrote the manuscript. Jordi Salmona, Lounès Chikhi, and Ute Radespiel contributed to the intellectual and

creative direction of the study. All authors read, revised, and approved the final manuscript.

## ACKNOWLEDGEMENTS

This is a Duke Lemur Centre publication #1510. We thank two anonymous reviewers and the editor for helpful comments that improved the manuscript.

## FUNDING INFORMATION

The research was supported by Duke Tropical Conservation Initiative Grant to ADY, CI/Primate Action Fund and Duke Lemur Center/SAVA Conservation research funds to MBB, Duke University research funds to ADY, a compute project grant provided by the North-German Supercomputing Alliance (HLRN) to UR and TvE, and the German Research Foundation (DFG Ra 502/23-1 to UR). LC and JS are supported by the LABEX entitled TULIP (ANR-10-LABX-41 and ANR-11-IDEX-0002-02) as well as the IRP BEEG-B (International Research Project-Bioinformatics, Ecology, Evolution, Genomics and Behaviour), and an "Investissement d'Avenir" grant (CEBA: ANR-10-LABX-25-01). LC is supported by the Fundação para a Ciência e Tecnologia (ref. PTDC-BIA-EVL/30815/2017). ADY would also like to acknowledge support from the John Simon Guggenheim Memorial Foundation and the Alexander von Humboldt Foundation.

## CONFLICT OF INTEREST

The authors declare no competing interests.

## DATA AVAILABILITY STATEMENT

All new sequencing data have been made available through NCBI BioProject PRJNA560399. Individual BioSample accessions are given in Table S1. VCFs, alignment files, phylogenies, and control files for demographic analyses will be available from the Dryad Digital Repository (<https://doi.org/10.5061/dryad.dncjsxkvz>). Additional scripts for data processing and analysis are available from TvE's GitHub page (<https://github.com/t-vane/ResearchSupplements>).

## ORCID

George P. Tiley  <https://orcid.org/0000-0003-0053-0207>

Tobias van Elst  <https://orcid.org/0000-0002-2601-6938>

Dominik Schüßler  <https://orcid.org/0000-0001-5885-7988>

Jordi Salmons  <https://orcid.org/0000-0002-1950-5805>

Amanda R. Stahlke  <https://orcid.org/0000-0001-5724-598X>

Paul A. Hohenlohe  <https://orcid.org/0000-0002-7616-0161>

Lounès Chikhi  <https://orcid.org/0000-0002-1140-0718>

Ute Radespiel  <https://orcid.org/0000-0002-0814-2404>

Anne D. Yoder  <https://orcid.org/0000-0002-1781-9552>

## REFERENCES

- Akaike, H. (1974). A new look at the statistical model identification. *IEEE Transactions on Automatic Control*, 19(6), 716–723. <https://doi.org/10.1109/TAC.1974.1100705>

- Ali, O. A., O'Rourke, S. M., Amish, S. J., Meek, M. H., Luikart, G., Jeffres, C., & Miller, M. R. (2016). Rad capture (Rapture): Flexible and efficient sequence-based genotyping. *Genetics*, 202(2), 389–400. <https://doi.org/10.1534/genetics.115.183665>
- Andriaholinirina, N., Baden, A., Blanco, M., Chikhi, L., Cooke, A., Davies, N., Dolch, R., Donati, G., Ganzhorn, J., Golden, C., Groeneveld, L. F., Hapke, A., Irwin, M., Johnson, S., Kappeler, P., King, T., Lewis, R., Louis, E. E., Markolf, M., ... Zaramody, A. (2014a). *Microcebus gerpi*. The IUCN Red List of Threatened Species. 2014:e.T16971461A16971464. <https://doi.org/10.2305/IUCN.UK.2014-1.RLTS.T16971461A16971464.en>
- Andriaholinirina, N., Baden, A., Blanco, M., Chikhi, L., Cooke, A., Davies, N., Dolch, R., Donati, G., Ganzhorn, J., Golden, C., Groeneveld, L. F., Hapke, A., Irwin, M., Johnson, S., Kappeler, P., King, T., Lewis, R., Louis, E. E., Markolf, M., ... Zaramody, A. (2014b). *Microcebus marohita*. The IUCN Red List of Threatened Species. 2014:e.T46251646A46251651. <https://doi.org/10.2305/IUCN.UK.2014-1.RLTS.T46251646A46251651.en>
- Andriambelason, J.-B., Blanco, M. B., Andriantsalohimisantatra, A., Rivoaharison, T. V., Walker, N., Birkinshaw, C., & Yoder, A. D. (2021). Living in tiny fragments: a glimpse at the ecology of Goodman's mouse lemurs (*Microcebus lehilahytsara*) in the relic forest of Ankafoke, Central Highlands, Madagascar. *Primates*, 62, 887–896. <https://doi.org/10.1007/s10329-021-00947-1>
- Blanco, M. B., Andriantsalohimisantatra, A. A., Rivoaharison, T. V., & Andriambelason, J. B. (2017). Evidence of prolonged torpor in Goodman's mouse lemurs at Ankafoke forest, central Madagascar. *Primates. Journal of Primatology*, 58(1), 31–37. <https://doi.org/10.1007/S10329-016-0586-3>
- Bolger, A. M., Lohse, M., & Usadel, B. (2014). Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics*, 30(15), 2114–2120. <https://doi.org/10.1093/bioinformatics/btu170>
- Bond, W. J., Silander, J. A., Jr., Ranaivonasy, J., & Ratirarson, J. (2008). The antiquity of Madagascar's grasslands and the rise of C<sub>4</sub> grassy biomes. *Journal of Biogeography*, 35, 1743–1758.
- Borgerson, C., Dolch, R., Ganzhorn, J., Greene, L. K., Guthrie, N. K., Johnson, S., Le Pors, B., Lewis, R., Louis, E. E., Rafalinirina, H. A., Raharivololona, B., Rakotoarisoa, G., Ralison, J., Randriahaingo, H. N. T., Rasoloarison, R. M., Raolinjanakolona, N. N., Sgarlata, G. M., Schwitzer, C., Wright, P. & Zaonarivelo, J. (2020). *Microcebus jollyae* (amended version of 2020 assessment). The IUCN Red List of Threatened Species 2020:e.T136458A180179376. <https://doi.org/10.2305/IUCN.UK.2020-3.RLTS.T136458A180179376.en>
- Bouckaert, R., Heled, J., Kühnert, D., Vaughan, T., Wu, C.-H., Xie, D., Suchard, M. A., Rambaut, A., & Drummond, A. J. (2014). BEAST 2: A software platform for Bayesian evolutionary analysis. *PLoS Computational Biology*, 10(4), e1003537. <https://doi.org/10.1371/journal.pcbi.1003537>
- Bowman, D. M. J. S., & Franklin, D. C. (2005). Fire ecology. *Progress in Physical Geography*, 29, 248–255.
- Bryant, D., Bouckaert, R., Felsenstein, J., Rosenberg, N. A., & RoyChoudhury, A. (2012). Inferring species trees directly from biallelic genetic markers: Bypassing gene trees in a full coalescent analysis. *Molecular Biology and Evolution*, 29(8), 1917–1932. <https://doi.org/10.1093/molbev>
- Burney, D. A. (1987a). Late Holocene vegetational change in central Madagascar. *Quaternary Research*, 28, 130–143.
- Burney, D. A. (1987b). Late Quaternary stratigraphic charcoal records from Madagascar. *Quaternary Research*, 28, 274–280.
- Burney, D. A. (1993). Late Holocene environmental changes in arid Southwestern Madagascar. *Quaternary Research*, 40, 98–106.
- Burney, D. A., Burney, L. P., Godfrey, L. R., Jungers, W. L., Goodman, S. M., Wright, H. T., & Jull, A. J. T. (2004). A chronology for late prehistoric Madagascar. *Journal of Human Evolution*, 47, 25–63.

- Burns, S. J., Godfrey, L. R., Faina, P., McGee, D., Hardt, B., Ranivoharimanana, L., & Randrianasy, J. (2016). Rapid human-induced landscape transformation in Madagascar at the end of the first millennium of the common era. *Quaternary Science Reviews*, 134, 92–99.
- Campbell, C. R., Tiley, G. P., Poelstra, J. W., Hunnicutt, K. E., Larsen, P. A., Lee, H.-J., Thorne, J. L., dos Reis, M., & Yoder, A. D. (2021). Substitution rate analysis validates surprising results from a pedigree-based measurement of the de novo mutation rate in gray mouse lemurs. *Heredity*, 127, 233–244. <https://doi.org/10.1038/s41437-021-0046-5>
- Chifman, J., & Kubatko, L. (2014). Quartet Inference from SNP data under the coalescent model. *Bioinformatics*, 30(23), 3317–3324. <https://doi.org/10.1093/bioinformatics/btu530>
- Chikhi, L., Sousa, V. C., Luisi, P., Goossens, B., & Beaumont, M. A. (2010). The confounding effects of population structure, genetic diversity and the sampling scheme on the detection and quantification of population size change. *Genetics*, 186, 983–995.
- Clark, P. U., Dyke, A. S., Shakun, J. D., Carlson, A. E., Clark, J., Wohlfarth, B., Mitrovica, J. X., Hostetler, S. W., & McCabe, A. M. (2009). The last glacial maximum. *Science*, 325, 710–714.
- Danecek, P., Auton, A., Abecasis, G., Albers, C. A., Banks, E., DePristo, M. A., Handsaker, R. E., Lunter, G., Marth, G. T., Sherry, S. T., McVean, G., Durbin, R., & 1000 Genomes Project Analysis Group. (2011). The variant call format and VCFtools. *Bioinformatics*, 27(15), 2156–2158. <https://doi.org/10.1093/BIOINFORMA/TICS/BTR330>
- Dewar, R. E., Radmilahy, C., Wright, H. T., Jacobs, Z., Kelly, G. O., & Berna, F. (2013). Stone tools and foraging in northern Madagascar challenge Holocene extinction models. *Proceedings of the National Academy of Sciences of the United States of America*, 110, 12583–12588.
- Dewar, R. E., & Wright, H. T. (1993). The culture history of Madagascar. *Journal of World Prehistory*, 7, 417–466.
- Dolch, R., Radespiel, U. & Blanco, M. (2020). *Microcebus lehilahytsara*. The IUCN Red List of Threatened Species 2020:e.T136199A115580879. <https://doi.org/10.2305/IUCN.UK.2020-2.RLTS.T136199A115580879.en>.
- Douglas, K., Hixon, S., Wright, H. T., Godfrey, L. R., Crowley, B. E., Manjakahery, B., Rasolondrainy, T., Crossland, Z., & Radmilahy, C. (2019). A critical review of radiocarbon dates clarifies the human settlement of Madagascar. *Quaternary Science Reviews*, 221, 105878. <https://doi.org/10.1016/j.quascirev.2019.105878>
- Edwards, E. J., & Smith, S. A. (2010). Phylogenetic analyses reveal the shady history of *C<sub>4</sub>* grasses. *Proceedings of the National Academy of Sciences of the United States of America*, 107, 2532–2537.
- Ersts, P. J. (2021). *Geographic distance matrix generator*. American Museum of Natural History, Center for Biodiversity and Conservation. [http://biodiversityinformatics.amnh.org/open\\_source/gdmg](http://biodiversityinformatics.amnh.org/open_source/gdmg)
- Etter, P. D., Bassham, S., Hohenlohe, P. A., Johnson, E. A., & Cresko, W. A. (2011). SNP discovery and genotyping for evolutionary genetics using RAD sequencing. *Methods in Molecular Biology*, 772, 157–178. [https://doi.org/10.1007/978-1-61779-228-1\\_9](https://doi.org/10.1007/978-1-61779-228-1_9)
- Evanno, G., Regnaut, S., & Goudet, J. (2005). Detecting the number of clusters of individuals using the software structure: A simulation study. *Molecular Ecology*, 14(8), 2611–2620. <https://doi.org/10.1111/J.1365-294X.2005.02553.X>
- Excoffier, L., Dupanloup, I., Huerta-Sánchez, E., Sousa, V. C., & Foll, M. (2013). Robust demographic inference from genomic and SNP data. *PLoS Genetics*, 9(10), e1003905. <https://doi.org/10.1371/journal.pgen.1003905>
- Excoffier, L., Smouse, P. E., & Quattro, J. M. (1992). Analysis of molecular variance inferred from metric distances among DNA haplotypes: Application to human mitochondrial DNA restriction data. *Genetics*, 131, 479–491.
- Farris, Z. J., Gerber, B. D., Valenta, K., Rafalirison, R., Razafimahaimodison, J. C., Larney, E., Rajaonarivelo, T., Randriana, Z., Wright, P. C., & Chapman, C. A. (2017). Threats to a rainforest carnivore community: A multi-year assessment of occupancy and co-occurrence in Madagascar. *Biological Conservation*, 210, 116–124.
- Felsenstein, J. (1981). Evolutionary trees from DNA sequences: A maximum likelihood approach. *Journal of Molecular Evolution*, 17, 368–376. <https://doi.org/10.1007/BF01734359>
- Foot, A. D., Vijay, N., Ávila-Arcos, M. C., Baird, R. W., Durban, J. W., Fumagalli, M., Gibbs, R. A., Hanson, M. B., Korneliusson, T. S., Martin, M. D., Robertson, K. M., Sousa, V. C., Vieira, F. G., Vinar, T., Wade, P., Worley, K. C., Excoffier, L., Morin, P. A., Gilbert, M. T. P., & Wolf, J. B. W. (2016). Genome-culture coevolution promotes rapid divergence of killer whale ecotypes. *Nature Communications*, 7, 11693. <https://doi.org/10.1038/ncomms11693>
- Gade, D. W. (1996). Deforestation and its effects in highland Madagascar. *Mountain Research and Development*, 16, 101–116.
- Gasse, F., & Van Campo, E. (1998). A 40,000-yr pollen and diatom record from Lake Tritrivakely, Madagascar, in the southern tropics. *Quaternary Research*, 49, 299–311.
- Genomic Resources Development Consortium, Blanchet, S., Bouchez, O., Chapman, C. A., Etter, P. D., Goldberg, T. L., Johnson, E. A., Jones, J. H., Loot, G., Omeja, P. A., Rey, O., Ruiz-Lopez, M. J., Switzer, W. M., & Ting, N. (2015). Genomic resources notes accepted 1 December 2014–31 January 2015. *Molecular Ecology Resources*, 15(3), 684. <https://doi.org/10.1111/1755-0998.12388>
- Gommery, D., Ramanivosoa, B., Faure, M., Guérin, C., Kerloc'h, P., Sénégas, F., & Randrianantenaina, H. (2011). Les plus anciennes traces d'activités anthropiques de Madagascar sur des ossements d'hippopotames subfossiles d'Anjohibe (Province de Mahajanga). *Comptes Rendus Palevol*, 10, 271–278.
- Goudet, J. (2005). hierfstat, a package for r to compute and test hierarchical F-statistics. *Molecular Ecology Notes*, 5(1), 184–186. <https://doi.org/10.1111/J.1471-8286.2004.00828.X>
- Green, G. M., & Sussman, R. W. (1990). Deforestation history of the eastern rain forests of Madagascar from satellite images. *Science*, 248(4952), 212–215. <https://doi.org/10.1126/science.248.4952.212>
- Hackel, J., Vorontsova, M. S., Nanjarisoa, O. P., Hall, R. C., Razanatsoa, J., Malakasi, P., & Besnard, G. (2018). Grass diversification in Madagascar: In situ radiation of two large *C<sub>3</sub>* shade clades and support for a Miocene to Pliocene origin of *C<sub>4</sub>* grassy biomes. *Journal of Biogeography*, 45(4), 750–761. <https://doi.org/10.1111/jbi.13147>
- Hansford, J., Wright, P. C., Rasoamiramanana, A., Pérez, V. R., Godfrey, L. R., Erickson, D., Thompson, T., & Turvey, S. T. (2018). Early Holocene human presence in Madagascar evidenced by exploitation of avian megafauna. *Science Advances*, 4, eaat6925.
- Harper, G. J., Steininger, M. K., Tucker, C. J., Juhn, D., & Hawkins, F. (2007). Fifty years of deforestation and forest fragmentation in Madagascar. *Environmental Conservation*, 34, 325–333.
- Heller, R., Nursyifa, C., Garcia Erill, G., Salmona, J., Chikhi, L., Meisner, J., Korneliusson, T. S., & Albrechtsen, A. (2021). A reference-free approach to analyze non-model RADseq data using standard Next Generation Sequencing toolkits. *Molecular Ecology Resources*, 21, 1085–1097. <https://doi.org/10.1111/1755-0998.13324>
- Humbert, H. (1927). La destruction d'une flore insulaire par le feu. Principaux aspects de la végétation à Madagascar. *Mémoires de L'Académie Malgache*, 5, 1–78.
- Jombart, T., & Ahmed, I. (2011). adegenet 1.3–1: New tools for the analysis of genome-wide SNP data. *Bioinformatics*, 27(21), 3070–3071. <https://doi.org/10.1093/bioinformatics/btr521>
- Joseph, G. S., Rakotoarivelo, A. R., & Seymour, C. L. (2021). How expansive were Malagasy Central Highland forests, ericoids, woodlands and grasslands? A multidisciplinary approach to a conservation conundrum. *Biological Conservation*, 261, 109282. <https://doi.org/10.1016/j.biocon.2021.109282>

- Kamvar, Z. N., Tabima, J. F., & Grünwald, N. J. (2014). Poppr: An R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ*, 2, e281. <https://doi.org/10.7717/peerj.281>
- Kass, R. E., & Raferty, A. E. (1995). Bayes factors. *Journal of the American Statistical Association*, 90, 773–795.
- Kishino, H., Miyata, T., & Hasegawa, M. (1990). Maximum likelihood inference of protein phylogeny and the origin of chloroplasts. *Journal of Molecular Evolution*, 31(2), 151–160. <https://doi.org/10.1007/BF02109483>
- Koechlin, J. (1993). Natural grasslands: eastern hemisphere and resume. In R. T. Coupland (Ed.), *Grasslands of Madagascar* (pp. 291–301). Elsevier.
- Korneliusson, T. S., Albrechtsen, A., & Nielsen, R. (2014). ANGSD: Analysis of next generation sequencing data. *BMC Bioinformatics*, 15(1), 356. <https://doi.org/10.1186/s12859-014-0356-4>
- Kozlov, A. M., Darriba, D., Flouri, T., Morel, B., & Stamatakis, A. (2019). RAxML-NG: A fast, scalable and user-friendly tool for maximum likelihood phylogenetic inference. *Bioinformatics*, 35(21), 4453–4455. <https://doi.org/10.1093/BIOINFORMATICS/BTZ305>
- Kull, C. A. (2003). Fire and the management of highland vegetation. In S. M. Goodman & J. P. Benstead (Eds.), *The natural history of Madagascar* (pp. 153–157). The University of Chicago Press.
- Kull, C. A. (2012). Air photo evidence of historical land cover change in the highlands: Wetlands and grasslands give way to crops and woodlots. *Madagascar Conservation & Development*, 7(3), 144–152.
- Lapierre, M., Lambert, A., & Achaz, G. (2017). Accuracy of demographic inferences from the site frequency spectrum: the case of the Yoruba population. *Genetics*, 206(1), 439–449. <https://doi.org/10.1534/genetics.116.192708>
- Larsen, P. A., Harris, R. A., Liu, Y., Murali, S. C., Campbell, C. R., Brown, A. D., Sullivan, B. A., Shelton, J., Brown, S. J., Raveendran, M., Dudchenko, O., Machol, I., Durand, N. C., Shamim, M. S., Aiden, E. L., Muzny, D. M., Gibbs, R. A., Yoder, A. D., Rogers, J., & Worley, K. C. (2017). Hybrid de novo genome assembly and centromere characterization of the gray mouse lemur (*Microcebus murinus*). *BMC Biology*, 15(1), 110. <https://doi.org/10.1186/s12915-017-0439-6>
- Lewis, P. O. (2001). A likelihood approach to estimating phylogeny from discrete morphological character data. *Systematic Biology*, 50, 913–925. <https://doi.org/10.1080/106351501753462876>
- Li, H. (2013). Aligning sequence reads, clone sequences and assembly contigs with BWA-MEM. *arXiv*, preprint, 1303.3997. <https://doi.org/10.48550/arXiv.1303.3997>
- Li, H., Handsaker, B., Wysoker, A., Fennell, T., Ruan, J., Homer, N., Marth, G., Abecasis, G., Durbin, R., & 1000 Genome Project Data Processing Subgroup. (2009). The Sequence Alignment/Map format and SAMtools. *Bioinformatics*, 25(16), 2078–2079. <https://doi.org/10.1093/bioinformatics/btp352>
- Liu, X., & Fu, Y.-X. (2015). Exploring population size changes using SNP frequency spectra. *Nature Genetics*, 47, 555–559.
- Liu, X., & Fu, Y.-X. (2020). Stairway Plot 2: demographic history inference with folded SNP frequency spectra. *Genome Biology*, 21, 280. <https://doi.org/10.1186/s13059-020-02196-9>
- Mazet, O., Rodríguez, W., Grusea, S., Boitard, S., & Chikhi, L. (2016). On the importance of being structured: instantaneous coalescence rates and human evolution – lessons for ancestral population size inference? *Heredity*, 116, 362–371.
- Meisner, J., & Albrechtsen, A. (2018). Inferring population structure and admixture proportions in low-depth NGS data. *Genetics*, 210(2), 719–731. <https://doi.org/10.1534/genetics.118.301336>
- Minh, B. Q., Schmidt, H. A., Chernomor, O., Schrempf, D., Woodhams, M. D., von Haeseler, A., & Lanfear, R. (2020). IQ-TREE 2: New models and efficient methods for phylogenetic inference in the genomic era. *Molecular Biology and Evolution*, 37(5), 1530–1534. <https://doi.org/10.1093/molbev/msaa015>
- Mitchell, P. (2019). Settling Madagascar: when did people first colonize the world's largest Island? *Journal of Island and Coastal Archaeology*, 15, 576–595. <https://doi.org/10.1080/15564894.2019.1582567>
- Muldoon, K. M., Crowley, B. E., Godfrey, L. R., Rasoamiramanana, A., Aronson, A., Jernvall, J., Wright, P. C., & Simons, E. L. (2012). Early Holocene fauna from a new subfossil site: A first assessment from Christmas River, south central Madagascar. *Madagascar Conservation and Development*, 7, 23–29.
- Myers, N., Mittermeier, R. A., da Fonseca, G. A. B., & Kent, J. (2000). Biodiversity hotspots for conservation priorities. *Nature*, 403(6772), 853–858. <https://doi.org/10.1038/35002501>
- Nielsen, R., Paul, J. S., Albrechtsen, A., & Song, Y. S. (2011). Genotype and ANP calling from next-generation sequencing data. *Nature Reviews. Genetics*, 12, 443–451. <https://doi.org/10.1038/nrg2986>
- Oksanen, J., Simpson, G. L., Blanchet, F. G., Kindt, R., Legendre, P., Minchin, P. R., O'Hara, R. B., Solymos, P., Stevens, M. H. H., Szoecs, E., Wagner, H., Barbour, M., Bedward, M., Bolker, B., Borcard, D., Carvalho, G., Chirico, M., de Caceres, M., Durand, S., ... Weedon, J. (2020). *vegan: Community ecology package*. <https://cran.r-project.org/package=vegan>
- O'Leary, S. J., Puritz, J. B., Willis, S. C., Hollenbeck, C. M., & Portnoy, D. S. (2018). These aren't the loci you're looking for: Principles of effective SNP filtering for molecular ecologists. *Molecular Ecology*, 27(16), 3193–3206. <https://doi.org/10.1111/mec.14792>
- Olivieri, G., Zimmermann, E., Randrianambinina, B., Rasoloharijaona, S., Rakotondravony, D., Guschanski, K., & Radespiel, U. (2007). The ever-increasing diversity in mouse lemurs: three new species in north and northwestern Madagascar. *Molecular Phylogenetics and Evolution*, 43(1), 309–327. <https://doi.org/10.1016/j.ympev.2006.10.026>
- Olivieri, G. L., Sousa, V., Chikhi, L., & Radespiel, U. (2008). From genetic diversity and structure to conservation: Genetic signature of recent population declines in three mouse lemur species (*Microcebus* spp.). *Biological Conservation*, 141, 1257–1271.
- Patton, A. H., Margres, M. J., Stahlke, A. R., Hendricks, S., Lewallen, K., Hamede, R. K., Ruiz-Aravena, M., Ryder, O., McCallum, H. I., Jones, M. E., Hohenlohe, P. A., & Storfer, A. (2019). Contemporary demographic reconstruction methods are robust to genome assembly quality: A case study in Tasmanian devils. *Molecular Biology and Evolution*, 36(12), 2906–2921. <https://doi.org/10.1093/molbev/msz191>
- Pedersen, C., Albrechtsen, A., Etter, P. D., Johnson, E. A., Orlando, L., Chikhi, L., Sigismund, H. R., & Heller, R. (2018). A southern African origin and cryptic structure in the highly mobile plains zebra. *Nature Ecology & Evolution*, 2, 491–498. <https://doi.org/10.1038/s41559-017-0453-7>
- Perrier de la Bâthie, H. (1921). La végétation Malgache. *Annales du Musée Colonial de Marseille*, 9, 1–226.
- Pierron, D., Heiske, M., Razafindrazaka, H., Rakoto, I., Rabetokotany, N., Ravololomanga, B., Rakotozafy, L. M.-A., Rakotomalala, M. M., Razafiarivony, M., Rasoarifetra, B., Raharijesy, M. A., Razafindralambo, L., Ramilisonina, F. F., Lejambale, S., Thomas, O., Abdallah, A. M., Rocher, C., Arachiche, A., ... Letellier, T. (2017). Genomic landscape of human diversity across Madagascar. *Proceedings of the National Academy of Sciences of the United States of America*, 114(32), E6498–E6506. <https://doi.org/10.1073/pnas.1704906114>
- Poelstra, J. W., Salmons, J., Tiley, G. P., Schübler, D., Blanco, M. B., Andriambeloso, J. B., Bouchez, O., Campbell, C. R., Etter, P. D., Hohenlohe, P. A., Hunnicutt, K. E., Iribar, A., Johnson, E. A., Kappeler, R., Larsen, P. A., Manzi, S., Ralison, J., Randrianambinina, B., Tasoliarison, R. M., ... Yoder, A. D. (2021). Cryptic patterns of speciation in cryptic primates: Microendemic mouse lemurs and the multispecies coalescent. *Systematic Biology*, 70(2), 203–218. <https://doi.org/10.1093/sysbio/syaa053>

- Pritchard, J. K., Stephens, M., & Donnelly, P. (2000). Inference of population structure using multilocus genotype data. *Genetics*, *155*, 945–959.
- Quéméré, E., Amelot, X., Pierson, J., Crouau-Roy, B., & Chikhi, L. (2012). Genetic data suggest a natural prehuman origin of open habitats in northern Madagascar and question the deforestation narrative in this region. *Proceedings of the National Academy of Sciences of the United States of America*, *109*, 13028–13033.
- Quéméré, E., Crouau-Roy, B., Rabarivola, C., Louis, E. E., Jr., & Chikhi, L. (2010). Landscape genetics of an endangered lemur (*Propithecus tattersalli*) within its entire fragmented range. *Molecular Ecology*, *19*, 1606–1621.
- Radespiel, U., Lutermann, H., Schmelting, B., & Zimmermann, E. (2019). An empirical estimate of the generation time of mouse lemurs. *American Journal of Primatology*, *81*(12), e23062. <https://doi.org/10.1002/ajp.23062>
- Radespiel, U., Ratsimbazafy, J. H., Rasoloharijaona, S., Raveloson, H., Andriaholinirina, N., Rakotondravony, R., Randrianarison, R. M., & Randrianambinina, B. (2012). First indications of a highland specialist among mouse lemurs (*Microcebus* spp.) and evidence for a new mouse lemur species from eastern Madagascar. *Primates*, *53*, 157–170.
- Railsback, L. B., Dupont, L. A., Liang, F., Brook, G. A., Burney, D. A., Cheng, H., & Edwards, R. L. (2020). Relationships between climate change, human environmental impact, and megafaunal extinction inferred from a 4000-year multi-proxy record from a stalagmite from northwestern Madagascar. *Quaternary Science Reviews*, *234*, 106244.
- Rambaut, A., Drummond, A. J., Xie, D., Baele, G., & Suchard, M. A. (2018). Posterior summarization in Bayesian phylogenetics using Tracer 1.7. *Systematic Biology*, *67*(5), 901–904. <https://doi.org/10.1093/sysbio/syy032>
- Rannala, B., & Yang, Z. (2003). Bayes estimation of species divergence times and ancestral population sizes using DNA sequences from multiple loci. *Genetics*, *164*(4), 1645–1656. <https://doi.org/10.1093/genetics/164.4.1645>
- Reynolds, J., Weir, B. S., & Cockerham, C. C. (1983). Estimation of the coancestry coefficient: basis for a short-term genetic distance. *Genetics*, *105*(3), 767–779. <https://doi.org/10.1093/genetics/105.3.767>
- Rochette, N. C., Rivera-Colón, A. G., & Catchen, J. M. (2019). Stacks 2: Analytical methods for paired-end sequencing improve RADseq-based population genomics. *Molecular Ecology*, *28*(21), 4737–4754. <https://doi.org/10.1111/mec.15253>
- Salmona, J., Heller, R., Quéméré, E., & Chikhi, L. (2017). Climate change and human colonization triggered habitat loss and fragmentation in Madagascar. *Molecular Ecology*, *26*, 5203–5222.
- Salmona, J., Olofsson, J. K., Hong-Wa, C., Razanatsoa, J., Rakotonasolo, F., Ralimanana, H., Randriamboavonjy, T., Suescun, U., Vorontsova, M. S., & Besnard, G. (2020). Late Miocene origin and recent population collapse of the Malagasy savanna olive tree. *Biological Journal of the Linnean Society*, *129*, 227–243.
- Samonds, K. E., Crowley, B. E., Rasolofomanana, T. R. N., Andriambelomanana, M. C., Andrianavalona, H. T., Ramihangihajason, T. N., & Godrey, L. R. (2019). A new late Pleistocene subfossil site (Tsaramody, Sambaina basin, central Madagascar) with implications for the chronology of habitat and megafaunal community change on Madagascar's Central Highlands. *Journal of Quaternary Science*, *34*, 379–392.
- Scholz, C. A., Cohen, A., Snsen, T. C., King, J., Talbot, M. R., & Brown, E. T. (2011). Scientific drilling in the great rift valley: The 2005 Lake Malawi scientific drilling project – an overview of the past 145,000 years of climate variability in southern hemisphere East Africa. *Palaeogeography, Palaeoclimatology, Palaeoecology*, *303*, 3–19.
- Schüßler, D., Blanco, M. B., Salmona, J., Poelstra, J., Andriambelason, J. B., Miller, A., Randrianambinina, B., Rasolofoson, D. W., Mantilla-Contreras, J., Chikhi, L., Louis, E. E., Jr., Yoder, A. D., & Radespiel, U. (2020). Ecology and morphology of mouse lemurs (*Microcebus* spp.) in a hotspot of microendemism in northeastern Madagascar, with the description of a new species. *American Journal of Primatology*, *82*(9), e23180. <https://doi.org/10.1002/ajp.23180>
- Schüßler, D., Mantilla-Contreras, J., Stadtmann, R., Ratsimbazafy, J. H., & Radespiel, U. (2020). Identification of crucial stepping stone habitats for biodiversity conservation in northeastern Madagascar using remote sensing and comparative predictive modeling. *Biodiversity and Conservation*, *29*(7), 2161–2184.
- Shimodaira, H. (2004). Approximately unbiased tests of regions using multistep-multiscale bootstrap resampling. *Annals of Statistics*, *32*, 2616–2641.
- Skotte, L., Korneliusen, T. S., & Albrechtsen, A. (2013). Estimating individual admixture proportions from next generation sequencing data. *Genetics*, *195*(3), 693–702. <https://doi.org/10.1534/genetics.113.154138>
- Solofondranohatra, C. L., Vorontsova, M. S., Hackel, J., Besnard, G., Cable, S., Williams, J., Jeannoda, V., & Lehmann, C. E. R. (2018). Grass functional traits differentiate forest and savanna in the Madagascar Central Highlands. *Frontiers in Ecology and Evolution*, *6*, 184.
- Styger, E., Rakotondramasy, H. M., Pfeffer, M. J., Fernandes, E. C. M., & Bates, D. M. (2007). Influence of slash-and-burn farming practices on fallow succession and land degradation in the rainforest region of Madagascar. *Agriculture, Ecosystems, and Environment*, *119*, 257–269.
- Swofford, D. L. (2003). *PAUP\*. Phylogenetic Analysis Using Parsimony (\*and Other Methods)*. Version 4. Sinauer Associates.
- Tavaré, S. (1986). Some probabilistic and statistical problems in the analysis of DNA sequences. *Lectures on Mathematics in the Life Sciences*, *17*, 57–86.
- Teixeira, H., Montade, V., Salmona, J., Metzger, J., Bremond, L., Kasper, T., Daut, G., Rouland, S., Ranarilalantiana, S., Rakotondravony, R., Chikhi, L., Behling, H., & Radespiel, U. (2021). Past environmental changes affected lemur population dynamics prior to human impact in Madagascar. *Communications Biology*, *4*, 1084. <https://doi.org/10.1038/s42003-021-02620-1>
- Teixeira, H., Salmona, J., Arredondo, A., Mourato, B., Manzi, S., Rakotondravony, R., Mazet, O., Chikhi, L., Metzger, J., & Radespiel, U. (2021). Impact of model assumptions on demographic inferences – The case study of two sympatric mouse lemurs in northwestern Madagascar. *BMC Ecology and Evolution*, *21*, 197. <https://doi.org/10.1186/s12862-021-01929-z>
- Vieilledent, G., Grinand, C., Rakotomalala, F. A., Ranaivosoa, R., Rakotoarijaona, J.-R., Allnutt, T. F., & Achard, F. (2018). Combining global tree cover loss data with historical national forest cover maps to look at six decades of deforestation and forest fragmentation in Madagascar. *Biological Conservation*, *222*, 189–197.
- Virah-Sawmy, M., Willis, K. J., & Gillson, L. (2009). Threshold response of Madagascar's littoral forest to sea-level rise. *Global Ecology and Biogeography*, *18*, 98–110.
- Virah-Sawmy, M., Willis, K. J., & Gillson, L. (2010). Evidence for drought and forest declines during the recent megafaunal extinctions in Madagascar. *Journal of Biogeography*, *37*, 506–519.
- Vorontsova, M. S., Besnard, G., Forest, F., Malakasi, P., Moat, J., Clayton, W. D., Ficinski, P., Savva, G. M., Nanjarisoa, O. P., Razanatsoa, J., Randriatsara, F. O., Kimeu, J. M., Luke, W. R. Q., Kayombo, C., & Linder, H. P. (2016). Madagascar's grasses and grasslands: anthropogenic or natural? *Proceedings of the Royal Society B: Biological Sciences*, *283*(1823), 20152226.
- Warmuth, V. M., & Ellegren, H. (2019). Genotype-free estimation of allele frequencies reduces bias and improves demographic inference

- from RADSeq data. *Molecular Ecology Resources*, 19(3), 586–596. <https://doi.org/10.1111/1755-0998.12990>
- Weir, B. S., & Cockerham, C. C. (1984). Estimating F-statistics for the analysis of population structure. *Evolution*, 38(6), 1358–1370. <https://doi.org/10.2307/2408641>
- Wilmé, L., Goodman, S. M., & Ganzhorn, J. U. (2006). Biogeographic evolution of Madagascar's microendemic biota. *Science*, 312, 1063–1065.
- Xie, W., Lewis, P. O., Fan, Y., Kuo, L., & Chen, M.-H. (2011). Improving marginal likelihood estimation for Bayesian phylogenetic model selection. *Systematic Biology*, 60, 150–160.
- Yang, Z. (1994). Maximum likelihood phylogenetic estimation from DNA sequences with variable rates over sites: Approximate methods. *Journal of Molecular Evolution*, 39(3), 306–314. <https://doi.org/10.1007/BF00160154>
- Yoder, A. D., Campbell, C. R., Blanco, M. B., dos Reis, M., Ganzhorn, J. U., Goodman, S. M., Hunnicutt, K. E., Larsen, P. A., KAppeler, P. M., Rasoloarison, R. M., Ralison, J. M., Swofford, D. L., & Weisrock, D. W. (2016). Geogenetic patterns in mouse lemurs (genus *Microcebus*) reveal the ghosts of Madagascar's forests past. *Proceedings of the National Academy of Sciences of the United States of America*, 113(29), 8049–8056. <https://doi.org/10.1073/pnas.1601081113>
- Zohdy, S., Gerber, B. D., Tecot, S., Blanco, M. B., Winshester, J. M., Wright, P. C., & Jernvall, J. (2014). Teeth, sex, and testosterone: Aging in the world's smallest primate. *PLoS One*, 9(10), e109528.

## SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

**How to cite this article:** Tiley, G. P., van Elst, T., Teixeira, H., Schüßler, D., Salmons, J., Blanco, M. B., Ralison, J. M., Randrianambinina, B., Rasoloarison, R. M., Stahlke, A. R., Hohenlohe, P. A., Chikhi, L., Louis, E. E., Radespiel, U., & Yoder, A. D. (2022). Population genomic structure in Goodman's mouse lemur reveals long-standing separation of Madagascar's Central Highlands and eastern rainforests. *Molecular Ecology*, 31, 4901–4918. <https://doi.org/10.1111/mec.16632>